

Supplementary Information:

Isolation of wheat bran-colonizing and metabolizing species from the human fecal microbiota

De Paepe et al., Submission to PeerJ

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Availability of raw data

This Supplementary Data file contains the R code to reproduce all analyses and plots presented in the manuscript entitled “Isolation of wheat bran-colonizing and metabolizing species from the human fecal microbiota”. All the required datasets are uploaded as additional supplementary files (DataS3-S9).

The raw 16S rRNA next-generation amplicon sequencing data is, moreover, deposited in the NCBI sequence read archive (SRA) under the accession number SRP091975 <https://trace.ncbi.nlm.nih.gov/Traces/study/?acc=SRP091975&go=go>. Raw 16S rRNA gene Sanger sequences of the isolates are supplied as a compressed folder (Sanger_isolates.zip). This folder and the raw Data files should be present in the same working directory containing this .Rmd script.

Load the packages and datasets

The following packages are required to run the script:

```
# Formatting data
library(reshape2)
library(plyr)
library(dplyr)
library(tidyr)
library(stringi)
library(stringr)
library(splitstackshape)
library(tibble)
library(gtools)

# Plotting data
library(ggplot2)
library(gridExtra)
```

```

library(cowplot)
require(scales)
library(gplots)

# User defined functions (make sure that the R scripts below are present in the
# same working directory as this Rmarkdown script)
source('DataS11.R') # formatting of ggplot graphs
source('DataS12.R') # formatting mothur taxonomy file

# Processing of 16S rRNA gene Sanger sequencing data
library("ape")
library("phangorn", lib.loc="/usr/local/lib/R/site-library")
source("https://bioconductor.org/biocLite.R")
biocLite("DECIPHER")
biocLite("Biostrings")
biocLite("sangerseqR")
library(devtools)
install_github("Liliad/sangeranalyseR")
library(sangeranalyseR)

# Processing of next-generation 16S rRNA gene amplicon sequencing data
biocLite("phyloseq")
library("phyloseq")
library(vegan)

# Rmarkdown knitr
library(knitr)

```

Load the functional data (short chain fatty acid production in the enrichment series):

```
enrichment_SCFA <- read.csv("DataS3.csv")
```

Load the 16S rRNA gene amplicon sequencing data (read count table and taxonomy):

```

# READ COUNT DATA
shared3 <- read.csv("DataS4.csv", header=TRUE)
rownames(shared3) <- shared3[,1]
shared3 <- shared3[, -1]

# Transpose
shared3 <- as.data.frame(t(shared3))

# Apply filtering cut-offs from the waste not want not paper by McMurdie and
# Holmes (2014)
shared3mingsingleton <- shared3[rowSums(shared3) != 0, ]
shared3mingsingleton <- shared3mingsingleton[rowSums(shared3mingsingleton) != 1, ]
minobs=1
prevalence <- apply(as.matrix(shared3mingsingleton), 1, function(x, minobs){sum(x
>=minobs)}, minobs)/ncol(shared3mingsingleton)
prevalencefilter <- prevalence > 0.05

```

```

shared3mingsingletonwnwn <- shared3mingsingleton[prevalencefilter,]
shared3mingsingletonwnwn <- shared3mingsingletonwnwn[ rowSums(shared3mingsingletonwnwn)>0.5*ncol(shared3mingsingletonwnwn),]

# Transform to proportions
shared3mingsingletonwnwnprop = sweep(shared3mingsingletonwnwn,2,colSums(shared3mingsingletonwnwn),'/')

#TAXONOMY
RDPTax <- read.csv("DataS6.csv")
RDPTax <- splittax(RDPTax,"otu")
rownames(RDPTax) <- RDPTax$id
RDPTax <- RDPTax[,-1]
RDPTax_noprob <- dplyr::select(RDPTax,-contains("prob"))

# Formatting
RDPTax_noprob$Genus <- mapvalues(RDPTax_noprob$Genus,from=c("Enterobacteriaceae_unclassified","Erysipelotrichaceae_unclassified","Lachnospiraceae_unclassified","Lactobacillales_unclassified","Actinobacteria_unclassified","Burkholderiales_unclassified","Clostridiales_unclassified","Ruminococcaceae_unclassified","Coriobacteriaceae_unclassified"),to=c("Enterobacteriaceae","Erysipelotrichaceae","Lachnospiraceae","Lactobacillales","Actinobacteria","Burkholderiales","Clostridiales","Ruminococcaceae","Coriobacteriaceae"))

#COMBINE IN OTU TABLE
otutab3 <- dplyr::select(RDPTax_noprob,-Species)
otutab3 <- cbind(otutab3[rownames(shared3mingsingletonwnwnprop),],shared3mingsingletonwnwnprop)

# Constructing Genus Level otutable with RDP taxonomy
genusotutab3 <- aggregate(. ~ Genus, data=otutab3[,-c(1:5)], FUN=sum)
rownames(genusotutab3) <- genusotutab3[,1]
genusotutab3 <- genusotutab3[,-1]

# Constructing Family Level otutable with with RDP taxonomy
familyotutab3 <- aggregate(. ~ Family, data=otutab3[,-c(1:4,6)], FUN=sum)
rownames(familyotutab3) <- familyotutab3[,1]
familyotutab3 <- familyotutab3[,-1]

# Constructing Order Level otutable with with RDP taxonomy
orderotutab3 <- aggregate(. ~ Order, data=otutab3[,-c(1:3,5,6)], FUN=sum)
rownames(orderotutab3) <- orderotutab3[,1]
orderotutab3 <- orderotutab3[,-1]

# Constructing Class Level otutable with with RDP taxonomy
classotutab3 <- aggregate(. ~ Class, data=otutab3[,-c(1:2,4:6)], FUN=sum)
rownames(classotutab3) <- classotutab3[,1]
classotutab3 <- classotutab3[,-1]

```

```

# Constructing Phylum Level otutable with with RDP taxonomy
phylumotutab3 <- aggregate(. ~ Phylum, data=otutab3[, -c(1,3:6)], FUN=sum)
rownames(phylumotutab3) <- phylumotutab3[,1]
phylumotutab3 <- phylumotutab3[, -1]

# ABUNDANCE DATA
shared4 <- read.csv("DataS5.csv", header=TRUE)
rownames(shared4) <- shared4[,1]
shared4 <- shared4[, -1]

# Transpose
shared4 <- as.data.frame(t(shared4))

# Apply filtering cut-offs from the waste not want not paper by McMurdie and
Holmes (2014)
shared4mingsingleton <- shared4[rowSums(shared4) != 0,]
shared4mingsingleton <- shared4mingsingleton[rowSums(shared4mingsingleton) != 1,]
minobs=1
prevalence <- apply(as.matrix(shared4mingsingleton), 1, function(x, minobs){sum(x
>=minobs)}, minobs)/ncol(shared4mingsingleton)
prevalencefilter <- prevalence > 0.05
shared4mingsingletonnwnwn <- shared4mingsingleton[prevalencefilter,]
shared4mingsingletonnwnwn <- shared4mingsingletonnwnwn[rowSums(shared4mingsingleto
nwnwn) > 0.5 * ncol(shared4mingsingletonnwnwn),]

# Transform to proportions
shared4mingsingletonnwnwnprop = sweep(shared4mingsingletonnwnwn, 2, colSums(shared4
mingsingletonnwnwn), '/')

# TAXONOMY DATA
RDPTax4 <- read.csv("DataS7.csv")
RDPTax4 <- splittax(RDPTax4, "otu")
rownames(RDPTax4) <- RDPTax4$id
RDPTax4 <- RDPTax4[, -1]
RDPTax4_noprob <- dplyr::select(RDPTax4, -contains("prob"))

RDPTax4_noprob$Genus <- mapvalues(RDPTax4_noprob$Genus, from=c("Enterobacteria
ceae_unclassified", "Erysipelotrichaceae_unclassified", "Lachnospiraceae_unclas
sified", "Lactobacillales_unclassified", "Actinobacteria_unclassified", "Burkhol
deriales_unclassified", "Clostridiales_unclassified", "Ruminococcaceae_unclassi
fied", "Coriobacteriaceae_unclassified", "Lactobacillaceae_unclassified", "Micro
bacteriaceae_unclassified"), to=c("Enterobacteriaceae", "Erysipelotrichaceae", "
Lachnospiraceae", "Lactobacillales", "Actinobacteria", "Burkholderiales", "Clostr
idiales", "Ruminococcaceae", "Coriobacteriaceae", "Lactobacillaceae", "Microbacte
riaceae"))

# COMBINE IN OTU TABLE
otutab4 <- dplyr::select(RDPTax4_noprob, -Species)
otutab4 <- cbind(otutab4[rownames(shared4mingsingletonnwnwnprop),], shared4minsi
ngletonnwnwnprop)

```

```

# Constructing Genus Level otutable with RDP taxonomy
genusotutab4 <- aggregate(. ~ Genus, data=otutab4[,-c(1:5)], FUN=sum)
rownames(genusotutab4) <- genusotutab4[,1]
genusotutab4 <- genusotutab4[,-1]

# Constructing Family Level otutable with with RDP taxonomy
familyotutab4 <- aggregate(. ~ Family, data=otutab4[,-c(1:4,6)], FUN=sum)
rownames(familyotutab4) <- familyotutab4[,1]
familyotutab4 <- familyotutab4[,-1]

# Constructing Order Level otutable with with RDP taxonomy
orderotutab4 <- aggregate(. ~ Order, data=otutab4[,-c(1:3,5,6)], FUN=sum)
rownames(orderotutab4) <- orderotutab4[,1]
orderotutab4 <- orderotutab4[,-1]

# Constructing Class Level otutable with with RDP taxonomy
classotutab4 <- aggregate(. ~ Class, data=otutab4[,-c(1:2,4:6)], FUN=sum)
rownames(classotutab4) <- classotutab4[,1]
classotutab4 <- classotutab4[,-1]

# Constructing Phylum Level otutable with with RDP taxonomy
phylumotutab4 <- aggregate(. ~ Phylum, data=otutab4[,-c(1,3:6)], FUN=sum)
rownames(phylumotutab4) <- phylumotutab4[,1]
phylumotutab4 <- phylumotutab4[,-1]

```

Load the metadata:

```

sampleinfo3 <- read.csv('DataS8.csv',header=TRUE)
sampleinfo3$Lumen_bran <- factor(sampleinfo3$Lumen_bran,levels=c("FS","Lumen",
" Bran"))
sampleinfo3$pH <-factor(sampleinfo3$pH,levels=c("5.8","6.8"))

sampleinfo4 <- read.csv('DataS9.csv',header=TRUE)
sampleinfo4$Lumen_bran <- factor(sampleinfo4$Lumen_bran,levels=c("FS","Lumen",
" Bran"))
sampleinfo4$pH <-factor(sampleinfo4$pH,levels=c("5.8","6.8"))

```

Specify color vectors:

```

# Colors
fourcolors <- c("#DC143C", "#8B5F65", "#00008B", "#EE9A00")
eightcolors=c("#7B4B71", "#C5CD42", "#C44F44", "#91B7C0", "#484934", "#BA9C5F", "#B
167C8", "#69C467")

elevencolors = c("#000000", "#CD6600", "#36648B", "#008B00", "#7A378B", "#838B83",
"#8B0000", "#323C4D", "#A03D44", "#98B736", "#C8ADA4")
fifteencolors <- c("#000000", "#CD6600", "#36648B", "#008B00", "#7A378B", "#838B83
", "#8B0000", "#323C4D", "#A03D44", "#98B736", "#C8ADA4", "#BB5236", "#7FCD50", "#AB5
CC7", "#C9A325")

```

```
thirtycolors=c("#323F24", "#CB51D7", "#72E245", "#DE4F2D", "#81DDC7", "#6584C6", "#CFAE3C", "#914261", "#BFDC86", "#CDC6BE", "#D3469A", "#3C315A", "#607B30", "#DD4469", "#5B9072", "#CA9FC7", "#7F592D", "#5A656D", "#D1867F", "#4C2426", "#79B6CE", "#6E67D1", "#CBDC3F", "#63D98B", "#97362B", "#CBB37D", "#7E3F85", "#CF8338", "#5DA73A", "#CB80D0")
```

```
# Resolution of figures
```

```
dpi <- 300
```

Sanger sequences of the isolates are supplied as a compressed folder (Sanger_isolates.zip).

SCFA production during consecutive enrichment steps

The fecal sample (enrichment step 0) and the liquid broth in the enrichment Hungate tubes after each transfer (enrichment steps 1 to 4) were sampled for SCFA analysis.

```
SCFA <- melt(enrichment_SCFA, id.vars=c("Donor", "pHstart", "Enrichment", "pHend"))
SCFA1 <- subset(SCFA, Donor=="1")
SCFA2 <- subset(SCFA, Donor=="2")
SCFA3 <- subset(SCFA, Donor=="3")
SCFA4 <- subset(SCFA, Donor=="4")
```

```
p <- ggplot(data=SCFA1, aes(x=Enrichment, y=as.numeric(as.character(value))))
p <- p + geom_bar(stat="identity", aes(fill=variable))
p <- papertheme(p) + facet_grid(pHstart~., drop=TRUE, scale="free_x", space="free_x") + scale_fill_manual(values=eightcolors, name=NULL)
p <- p + ylab("Donor 1 SCFA (mM)") + xlab("Enrichment step") + theme(strip.text.y = element_text(angle=0), legend.justification="center") + guides(fill=guide_legend(nrow=2))
p <- p + geom_text(data=SCFA1, aes(x=Enrichment, y=100, label=pHend), size=6)
mylegendSCFA <- get_legend(p)
SCFAAd1 <- p + theme(legend.position = "none", axis.title.x = element_text(color="white"))
```

```
p <- ggplot(data=SCFA1, aes(x=Enrichment, y=as.numeric(as.character(value))))
p <- p + geom_bar(stat="identity", aes(fill=variable))
p <- papertheme(p) + facet_grid(pHstart~., drop=TRUE, scale="free_x", space="free_x") + scale_fill_manual(values=eightcolors, name=NULL)
p <- p + ylab("Donor 1 SCFA (mM)") + xlab("Enrichment step") + theme(strip.text.y = element_text(angle=0), legend.justification="center") + guides(fill=guide_legend(nrow=2))
p <- p + geom_text(data=SCFA1, aes(x=Enrichment, y=100, label=pHend), size=6)
mylegendSCFA <- get_legend(p)
SCFAAd1 <- p + theme(legend.position = "none", axis.title.x = element_text(color="white"))
```

```
p <- ggplot(data=SCFA2, aes(x=Enrichment, y=as.numeric(as.character(value))))
p <- p + geom_bar(stat="identity", aes(fill=variable))
```

```

p <- papertheme(p) +facet_grid(pHstart~.,drop=TRUE,scale="free_x",space="free_x") + scale_fill_manual(values=eightcolors)
p <- p+ ylab("Donor 2 SCFA (mM)") + xlab("Enrichment step") + theme(legend.text=element_text(face='italic'),strip.text.y = element_text(angle=0)) +guides(fill=guide_legend(nrow=4))
p <- p + geom_text(data=SCFA2,aes(x=Enrichment,y=85,label=pHend),size=6)
SCFAd2 <- p + theme(legend.position = "none",axis.title.x = element_text(color="white"))

p <- ggplot(data=SCFA3,aes(x=Enrichment,y=as.numeric(as.character(value))))
p <- p +geom_bar(stat="identity",aes(fill=variable))
p <- papertheme(p) +facet_grid(pHstart~.,drop=TRUE,scale="free_x",space="free_x") + scale_fill_manual(values=eightcolors)
p <- p+ ylab("Donor 3 SCFA (mM)") + xlab("Enrichment step") + theme(legend.text=element_text(face='italic'),strip.text.y = element_text(angle=0)) +guides(fill=guide_legend(nrow=4))
p <- p + geom_text(data=SCFA3,aes(x=Enrichment,y=100,label=pHend),size=6)
SCFAd3 <- p + theme(legend.position = "none")

p <- ggplot(data=SCFA4,aes(x=Enrichment,y=as.numeric(as.character(value))))
p <- p +geom_bar(stat="identity",aes(fill=variable))
p <- papertheme(p) +facet_grid(pHstart~.,drop=TRUE,scale="free_x",space="free_x") + scale_fill_manual(values=eightcolors)
p <- p+ ylab("Donor 4 SCFA (mM)") + xlab("Enrichment step") + theme(legend.text=element_text(face='italic'),strip.text.y = element_text(angle=0)) +guides(fill=guide_legend(nrow=4))
p <- p + geom_text(data=SCFA4,aes(x=Enrichment,y=100,label=pHend),size=6)
SCFAd4 <- p + theme(legend.position = "none")

tiff("Figure2.tiff",width=10*dpi,height=16*dpi,res=dpi,compression="lzw")
cowplot::plot_grid(plot_grid(SCFAd1,SCFAd2,SCFAd3,SCFAd4,nrow=2,ncol=2,rel_heights =c(0.5,0.5,0.5,0.5),rel_widths=c(0.5,0.5,0.5,0.5),labels='AUTO',label_size=20,label_fontfamily='sans'),mylegendSCFA,nrow=2,rel_heights=c(0.95,0.05))
dev.off()

## png
## 2

knitr::include_graphics("Figure2.tiff")

```

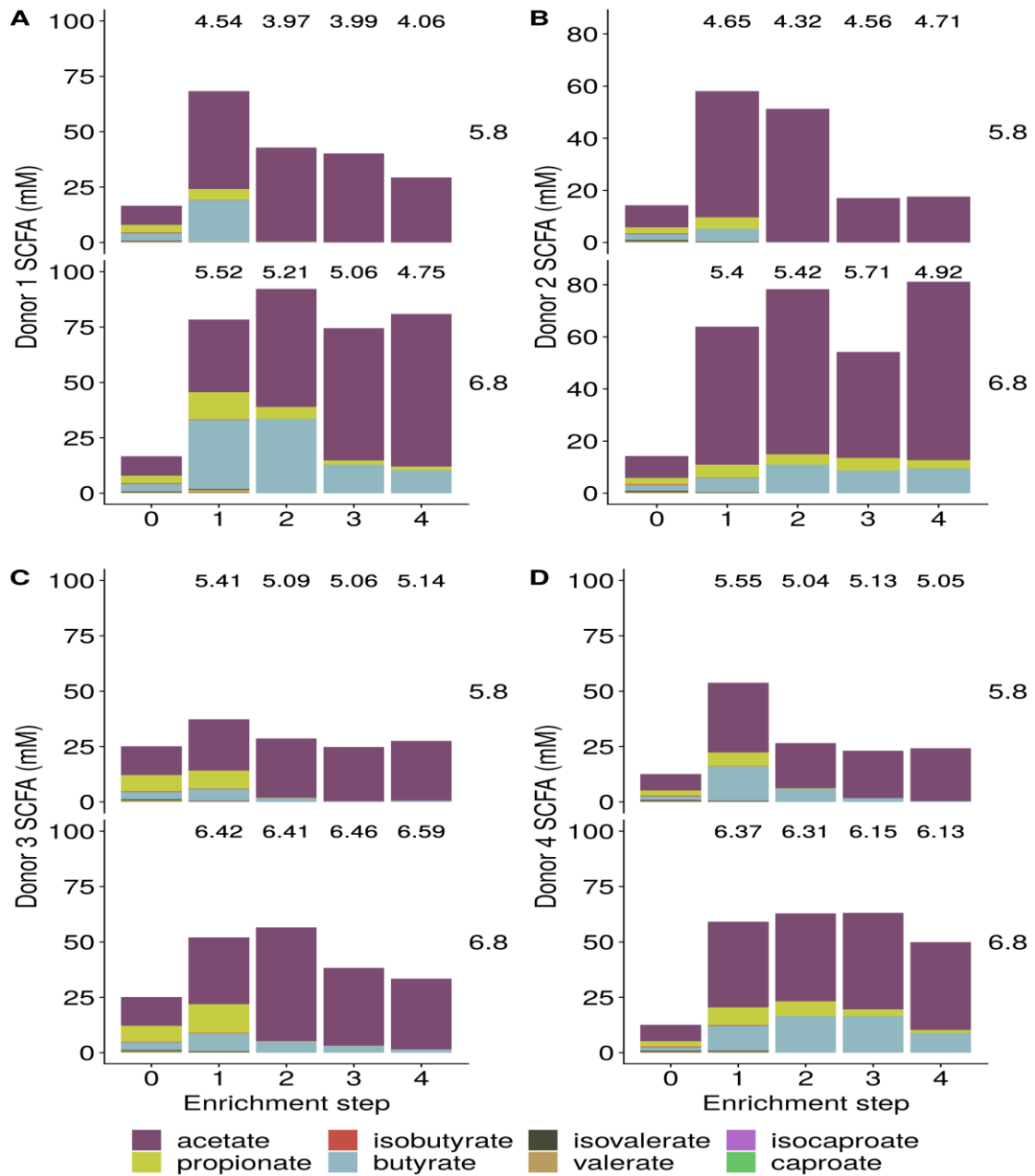


Figure 2: Short Chain Fatty Acid((SCFA) production by the fecal microbiota derived from four different donors during enrichment with wheat bran as the sole nutrient source. The fecal sample (enrichment step 0) was incubated with wheat bran for 24 h (enrichment step 1), after which the wheat bran residue was washed to remove loosely attached bacteria and used to seed a new incubation (enrichment step 2). This procedure was repeated two more times (enrichment step 3 and 4). The pH is indicated on top of the stacked bars and decreased considerably comparing to the starting pH (5.8 and 6.8).

Richness and Diversity indices calculated on the next-generation 16S rRNA gene amplicon sequencing confirm the 'enrichment'

The outcome of the enrichment procedure was assessed by computing richness (Chao1 Richness estimator) and diversity (Shannon, Simpson, inverse Simpson and Fisher alpha) estimators using `vegan_2.4-4` (Oksanen et al. 2016).

```
dataunnormalised <- as.data.frame(t(shared3))
# Calculate the chao index
chaoest=apply(dataunnormalised,1,function(x) estimateR(x))[2,]
# Annotate the dataframe
richnessdf <- cbind(chaoest,sampleinfo3)
richnessdf$Enrichment <-as.numeric(as.character(richnessdf$Enrichment))
richness1 <- subset(richnessdf,Donor=="1"&Nameshared!="Donor2_low4_Brep.V4")
richness2 <- subset(richnessdf,Donor=="2")
richness3<- subset(richnessdf,Donor=="3")
# Repeat for donor 4
dataunnormalised <- as.data.frame(t(shared4))
chaoest=apply(dataunnormalised,1,function(x) estimateR(x))[2,]
richnessdf4 <- cbind(chaoest,sampleinfo4)
richnessdf4$Enrichment <- as.numeric(as.character(richnessdf4$Enrichment))
richness4 <- subset(richnessdf4,Donor=="4")

# Evolution of the chao richness during consecutive enrichment steps in donor
1 (Figure 3A in the manuscript)
p <- ggplot(data=richness1,aes(x=Enrichment,y=as.numeric(as.character(chaoest
))))
p <- p +geom_line(aes(group=Lumen_bran,linetype=Lumen_bran))+geom_point(aes(s
hape=Lumen_bran),size=5)
p <- papertheme(p) +facet_grid(pH~.,drop=TRUE,scale="free_x",space="free_x")
+ scale_fill_manual(values=eightcolors,name=NULL) + scale_shape_manual(name=N
ULL,values=c(15,16,17))+ scale_linetype_manual(name=NULL,values=c(0,1,2))
p <- p+ ylab("Chao1 richness estimator \n Donor1") + xlab("Enrichment step")
+ theme(strip.text.y = element_text(angle=0)) +guides(fill=guide_legend(nrow=
2))+ theme(legend.key.width = unit(2,"cm"))
richplot1 <- p + theme(legend.position = "none",axis.title.x = element_text(c
olor="white"))

# Evolution of the chao richness during consecutive enrichment steps in three
donors
richness <- rbind(richness1,richness2,richness3)
richness$Donor <- as.factor(richness$Donor)

p <- ggplot(data=richness,aes(x=Enrichment,y=as.numeric(as.character(chaoest
))))
p <- p +geom_line(aes(group=interaction(Lumen_bran,Donor),color=Donor))+geom_
point(aes(shape=Lumen_bran,color=Donor),size=5)
p <- papertheme(p) +facet_grid(pH~.,drop=TRUE,scale="free_x",space="free_x")
+ scale_color_manual(values=fourcolors,name=NULL)
```

```

p <- p + ylab("Chao richness estimator") + xlab("Enrichment step") + theme(leg
end.text=element_text(face='italic'),strip.text.y = element_text(angle=0)) +g
uides(fill=guide_legend(nrow=2))
richplot <- p + theme(legend.position = "none",axis.title.x = element_text(co
lor="white"))

dataunnormalised <- as.data.frame(t(shared3))
# Calculate different diversity indices
divdf <- data.frame(shannon=apply(dataunnormalised,1,function(x) vegan::diver
sity(x)),simpson=apply(dataunnormalised,1,function(x) vegan::diversity(x,"sim
p")),invsimpson=apply(dataunnormalised,1,function(x) vegan::diversity(x,"inv"
)),fisher=apply(dataunnormalised,1,function(x) vegan::fisher.alpha(x)))
# Annotate the dataframe
divdf <- cbind(divdf,sampleinfo3)
divdf$Enrichment <- as.numeric(as.character(divdf$Enrichment))
divdf1 <- subset(divdf,Donor=="1"&Nameshared!="Donor2_low4_Brep.V4")
divdf1$Lumen_bran <- mapvalues(divdf1$Lumen_bran,from="FS",to="Fecal sample")

divdf2 <- subset(divdf,Donor=="2")
divdf3<- subset(divdf,Donor=="3")
# Repeat for donor 4
dataunnormalised <- as.data.frame(t(shared4))
divdf4 <- data.frame(shannon=apply(dataunnormalised,1,function(x) vegan::dive
rsity(x)),simpson=apply(dataunnormalised,1,function(x) vegan::diversity(x,"si
mp")),invsimpson=apply(dataunnormalised,1,function(x) vegan::diversity(x,"inv
")),fisher=apply(dataunnormalised,1,function(x) vegan::fisher.alpha(x)))
divdf4 <- cbind(divdf4,sampleinfo4)

# Evolution of the diversity indices during consecutive enrichment steps in d
onor 1 (Figure 3B in the manuscript)
p <- ggplot(data=divdf1,aes(x=Enrichment,y=as.numeric(as.character(shannon)))
)
p <- p +geom_line(aes(group=Lumen_bran,linetype=Lumen_bran))+geom_point(aes(s
hape=Lumen_bran),size=5)
p <- papertHEME(p) +facet_grid(pH~.,drop=TRUE,scale="free_x",space="free_x")
+ scale_fill_manual(values=eightcolors,name=NULL) + scale_shape_manual(name=N
ULL,values=c(15,16,17)) + scale_linetype_manual(name=NULL,values=c(0,1,2))
p <- p+ ylab("Shannon diversity index \n Donor 1") + xlab("Enrichment step")
+ theme(strip.text.y = element_text(angle=0)) +guides(fill=guide_legend(nrow=
2)) + theme(legend.justification="center") + theme(legend.key.width = unit(2,
"cm"))
mylegendrichndiv <- get_legend(p)
divplot1 <- p + theme(legend.position = "none")
# Evolution of the diversity indices during consecutive enrichment steps thre
e donors
divdf <- rbind(divdf1,divdf2,divdf3)
divdf$Donor <- as.factor(divdf$Donor)

p <- ggplot(data=divdf,aes(x=Enrichment,y=as.numeric(as.character(shannon))))
p <- p +geom_line(aes(group=interaction(Lumen_bran,Donor),color=Donor))+geom_

```

```

point(aes(shape=Lumen_bran,color=Donor),size=5)
p <- papertheme(p) +facet_grid(pH~.,drop=TRUE,scale="free_x",space="free_x")
+ scale_color_manual(values=fourcolors,name=NULL)
p <- p+ ylab("Chao richness estimator") + xlab("Enrichment step") + theme(leg
end.text=element_text(face='italic'),strip.text.y = element_text(angle=0)) +g
uides(fill=guide_legend(nrow=2))
p <- p + theme(legend.position = "none",axis.title.x = element_text(color="wh
ite"))

tiff("Figure3.tiff",width=6*dpi,height=14*dpi,res=dpi,compression="lzw")
cowplot::plot_grid(plot_grid(richplot1,divplot1,ncol=1,rel_heights = c(0.5,0.
5),labels='AUTO',label_size=20,label_fontfamily='sans'),mylegendrichndiv,nrow
=2,rel_heights=c(0.95,0.05))
dev.off()

## png
## 2

include_graphics("Figure3.tiff")

```

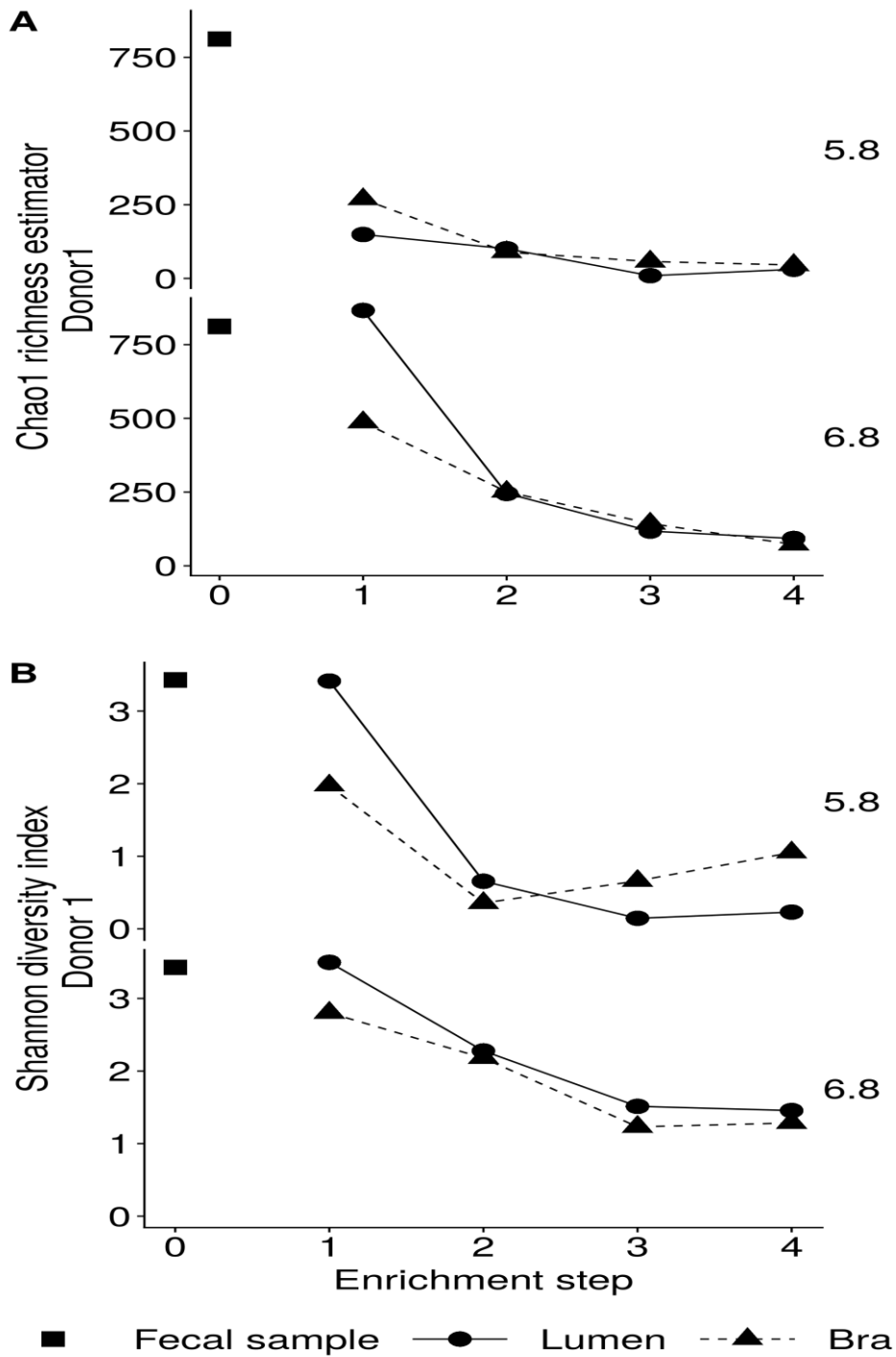


Figure 3: Reduction in the microbial community richness (Chao 1 richness estimator) and diversity (Shannon diversity index) during consecutive enrichment steps with wheat bran as the sole nutrient source, as shown for donor 1. The fecal sample (enrichment step 0) was incubated with wheat bran for 24 h (enrichment step 1), after which the wheat bran residue was washed to remove loosely attached bacteria and used to seed a new incubation (enrichment step 2). This procedure was repeated two more times (enrichment step 3 and 4).

Identification of the enriched species by next-generation 16S rRNA gene amplicon sequencing

Bar graphs of the proportional microbial community composition at phylum, genus, and species level were created to demonstrate community shifts during consecutive enrichment steps with wheat bran as the sole nutrient source.

```
otuspec <- phylumotutab3
otuspec <- 100*otuspec
otuspec <- rbind(names=colnames(otuspec),otuspec)
otuspec <- mutate(as.data.frame(t(otuspec)),donor=sampleinfo3$Donor,LBM=sampleinfo3$Lumen_bran,enrichment=sampleinfo3$Enrichment,pH=sampleinfo3$pH)
otuspec <- melt(otuspec,id.vars=c('donor','LBM','enrichment','pH','names'))
otuspec1 <- subset(otuspec,donor=="1"&enrichment!="c"&names!="Donor2_low4_Brep.V4")
otuspec2 <- subset(otuspec,donor=="2"&enrichment!="c"&names!="Donor2_low4_Brep.V4")
otuspec3<- subset(otuspec,donor=="3"&enrichment!="c"&names!="Donor2_low4_Brep.V4")

# Donor1
p <- ggplot(data=otuspec1,aes(x=LBM,y=as.numeric(as.character(value))))
p <- p +geom_bar(stat="identity",aes(fill=variable))
p <- papertheme(p) +facet_grid(pH~enrichment,drop=TRUE,scale="free_x",space="free_x") + scale_fill_manual(values=elevencolors)
p <- p+ ylab("Relative abundance (%)") + xlab(NULL) + theme(legend.text=element_text(face='italic'),strip.text.y = element_text(angle=0)) +guides(fill=guide_legend(nrow=4))

# Donor2
p <- ggplot(data=otuspec2,aes(x=LBM,y=as.numeric(as.character(value))))
p <- p +geom_bar(stat="identity",aes(fill=variable))
p <- papertheme(p) +facet_grid(pH~enrichment,drop=TRUE,scale="free_x",space="free_x") + scale_fill_manual(values=elevencolors)
p <- p+ ylab("Relative abundance (%)") + xlab(NULL) + theme(legend.text=element_text(face='italic'),strip.text.y = element_text(angle=0)) +guides(fill=guide_legend(nrow=4))

#Donor3
p <- ggplot(data=otuspec3,aes(x=LBM,y=as.numeric(as.character(value))))
p <- p +geom_bar(stat="identity",aes(fill=variable))
p <- papertheme(p) +facet_grid(pH~enrichment,drop=TRUE,scale="free_x",space="free_x") + scale_fill_manual(values=elevencolors)
p <- p+ ylab("Relative abundance (%)") + xlab(NULL) + theme(legend.text=element_text(face='italic'),strip.text.y = element_text(angle=0)) +guides(fill=guide_legend(nrow=4))

otuspec<- genusotutab3
otuspec <- cbind(sampleinfo3,t(otuspec))
```

```

otuspec <- subset(otuspec, Enrichment!="c")
otuspec <- otuspec[, -c(1:length(colnames(sampleinfo3)))]
otuspec <- data.frame(t(otuspec))
selecttop20par <- rowSums(otuspec)
otuspec <- tibble::rownames_to_column(otuspec)
otuspec <- as.data.frame(otuspec %>% top_n(13, selecttop20par))
other <- 1-colSums(otuspec[, -1])
rownames(otuspec) <- otuspec$rowname
otuspec <- otuspec[, -1]
otuspec <- rbind(otuspec, other)
rownames(otuspec)[nrow(otuspec)] <- "Other"
otuspec <- 100*otuspec

sampleinfosub3 <- subset(sampleinfo3, Enrichment!="c")
otuspec <- mutate(as.data.frame(t(otuspec)), donor=sampleinfosub3$Donor, LBM=sampleinfosub3$Lumen_bran, enrichment=sampleinfosub3$Enrichment, pH=sampleinfosub3$pH, names=colnames(otuspec))
otuspec <- melt(otuspec, id.vars=c('donor', 'LBM', 'enrichment', 'pH', 'names'))
otuspec1 <- subset(otuspec, donor=="1"&names!="Donor2_low4_Brep.V4")
otuspec2 <- subset(otuspec, donor=="2"&names!="Donor2_low4_Brep.V4")
otuspec3 <- subset(otuspec, donor=="3"&names!="Donor2_low4_Brep.V4")

# Donor1
p <- ggplot(data=otuspec1, aes(x=LBM, y=as.numeric(as.character(value))))
p <- p +geom_bar(stat="identity", aes(fill=variable))
p <- papertheme(p) +facet_grid(pH~enrichment, drop=TRUE, scale="free_x", space="free_x") + scale_fill_manual(values=fifteencolors, name="Genus")
p <- p+ ylab("Relative abundance (%)") + xlab(NULL) + theme(legend.text=element_text(face='italic'), strip.text.y = element_text(angle=0), legend.position="right") +guides(fill=guide_legend(ncol=1))
bargraph1genus <- p

# Donor2
p <- ggplot(data=otuspec2, aes(x=LBM, y=as.numeric(as.character(value))))
p <- p +geom_bar(stat="identity", aes(fill=variable))
p <- papertheme(p) +facet_grid(pH~enrichment, drop=TRUE, scale="free_x", space="free_x") + scale_fill_manual(values=fifteencolors, name="Genus")
p <- p+ ylab("Relative abundance (%)") + xlab(NULL) + theme(legend.text=element_text(face='italic'), strip.text.y = element_text(angle=0), legend.position="right") +guides(fill=guide_legend(ncol=1))

# Donor3
p <- ggplot(data=otuspec3, aes(x=LBM, y=as.numeric(as.character(value))))
p <- p +geom_bar(stat="identity", aes(fill=variable))
p <- papertheme(p) +facet_grid(pH~enrichment, drop=TRUE, scale="free_x", space="free_x") + scale_fill_manual(values=fifteencolors, name="Genus")
p <- p+ ylab("Relative abundance (%)") + xlab(NULL) + theme(legend.text=element_text(face='italic'), strip.text.y = element_text(angle=0), legend.position="right") +guides(fill=guide_legend(ncol=1))

```

```

# Donor4
otuspec<- genusotutab4
selecttop20par <- rowSums(otuspec)
otuspec <- tibble::rownames_to_column(otuspec)
otuspec <- as.data.frame(otuspec %>% top_n(13,selecttop20par))
other <- 1-colSums(otuspec[,-1])
rownames(otuspec) <- otuspec$rowname
otuspec <- otuspec[,-1]
otuspec <- rbind(otuspec,other)
rownames(otuspec)[nrow(otuspec)] <- "Other"
otuspec <- otuspec*100

otuspec <- mutate(as.data.frame(t(otuspec)),donor=sampleinfo4$Donor,LBM=sampleinfo4$Lumen_bran,enrichment=sampleinfo4$Enrichment,pH=sampleinfo4$pH, names=colnames(otuspec))
otuspec4 <- melt(otuspec,id.vars=c('donor','LBM','enrichment','pH','names'))

p <- ggplot(data=otuspec4,aes(x=LBM,y=as.numeric(as.character(value))))
p <- p +geom_bar(stat="identity",aes(fill=variable))
p <- papertheme(p) +facet_grid(pH~enrichment,drop=TRUE,scale="free_x",space="free_x") + scale_fill_manual(values=fifteencolors,name="Genus")
p <- p+ ylab("Relative abundance (%)") + xlab(NULL) + theme(legend.text=element_text(face='italic'),strip.text.y = element_text(angle=0),legend.position="right") +guides(fill=guide_legend(ncol=1))

# Species level annotation of OTUs (verified by BLAST and RDP)
label3donorsillu <- c("OTU1 Bifidobacterium sp.,"OTU2 Enterobacteriaceae sp.",
,"OTU3 Escherichia/Shigella sp.,"OTU4 Lactobacillus sp.,"OTU5 Faecalibacterium prausnitzii",
"OTU6 Prevotella copri","OTU7 Faecalibacterium prausnitzii",
"OTU8 Pediococcus sp.,"OTU9 E. rectale/R. faecis","OTU10 Fusobacterium mortiferum",
"OTU11 Bacteroides dorei","OTU12 Dialister sp.,"OTU13 Sutterella wadsworthensis",
"OTU14 Lactobacillus sp.,"OTU15","OTU16 Bacteroides eggerthii",
"OTU17 Blautia wexlerae","OTU18 Lactobacillus fermentus","OTU19 Fusicatenibacter saccharivorans",
"OTU20 Roseburia faecis","Other")
otuspec<-otutab3[,-c(1:6)]
otuspec <- cbind(sampleinfo3,t(otuspec))
otuspec <- subset(otuspec,Enrichment!="c")
otuspec <- otuspec[,-c(1:length(colnames(sampleinfo3)))]
otuspec <- data.frame(t(otuspec))
selecttop20par <- rowSums(otuspec)
otuspec <- tibble::rownames_to_column(otuspec)
otuspec <- as.data.frame(otuspec %>% top_n(20,selecttop20par))
other <- 1-colSums(otuspec[,-1])
rownames(otuspec) <- otuspec$rowname
otuspec <- otuspec[,-1]
otuspec <- rbind(otuspec,other)
rownames(otuspec)[nrow(otuspec)] <- "Other"
otuspec <- 100*otuspec

sampleinfosub3 <- subset(sampleinfo3,Enrichment!="c")

```



```

otuspec <- mutate(as.data.frame(t(otuspec)),donor=sampleinfosub3$Donor,LBM=sa
mpleinfosub3$Lumen_bran,enrichment=sampleinfosub3$Enrichment,pH=sampleinfosub
3$pH,names=colnames(otuspec))
otuspec <- melt(otuspec,id.vars=c('donor','LBM','enrichment','pH','names'))
otuspec1 <- subset(otuspec,donor=="1"&names!="Donor2_low4_Brep.V4")
otuspec2 <- subset(otuspec,donor=="2"&names!="Donor2_low4_Brep.V4")
otuspec3<- subset(otuspec,donor=="3"&names!="Donor2_low4_Brep.V4")

# Donor1
p <- ggplot(data=otuspec1,aes(x=LBM,y=as.numeric(as.character(value))))
p <- p +geom_bar(stat="identity",aes(fill=variable))
p <- papertheme(p) +facet_grid(pH~enrichment,drop=TRUE,scale="free_x",space="
free_x") + scale_fill_manual(values=thirtycolors,name="Species",labels=label3
donorsillu)
p <- p+ ylab("Relative abundance (%)") + xlab(NULL) + theme(legend.text=elemen
t_text(face='italic'),strip.text.y = element_text(angle=0),legend.position="
right") +guides(fill=guide_legend(ncol=1))
bargraph1species <- p

#Donor2
p <- ggplot(data=otuspec2,aes(x=LBM,y=as.numeric(as.character(value))))
p <- p +geom_bar(stat="identity",aes(fill=variable))
p <- papertheme(p) +facet_grid(pH~enrichment,drop=TRUE,scale="free_x",space="
free_x") + scale_fill_manual(values=thirtycolors,name="Species")
p <- p+ ylab("Relative abundance (%)") + xlab(NULL) + theme(legend.text=elemen
t_text(face='italic'),legend.position="right",strip.text.y = element_text(an
gle=0)) +guides(fill=guide_legend(ncol=1))

# Donor3
p <- ggplot(data=otuspec3,aes(x=LBM,y=as.numeric(as.character(value))))
p <- p +geom_bar(stat="identity",aes(fill=variable))
p <- papertheme(p) +facet_grid(pH~enrichment,drop=TRUE,scale="free_x",space="
free_x") + scale_fill_manual(values=thirtycolors,name="Species")
p <- p+ ylab("Relative abundance (%)") + xlab(NULL) + theme(legend.text=elemen
t_text(face='italic'),strip.text.y = element_text(angle=0),legend.position="
right") +guides(fill=guide_legend(ncol=1))

tiff("Figure4.tiff",width=16*dpi,height=14*dpi,res=dpi,compression="lzw")
cowplot::plot_grid(bargraph1genus,bargraph1species,nrow=2,rel_heights =c(0.5,
0.5),labels='AUTO',label_size=20,label_fontfamily='sans',align="v",axis="1")
dev.off()

## png
## 2

knitr::include_graphics("Figure4.tiff")

```

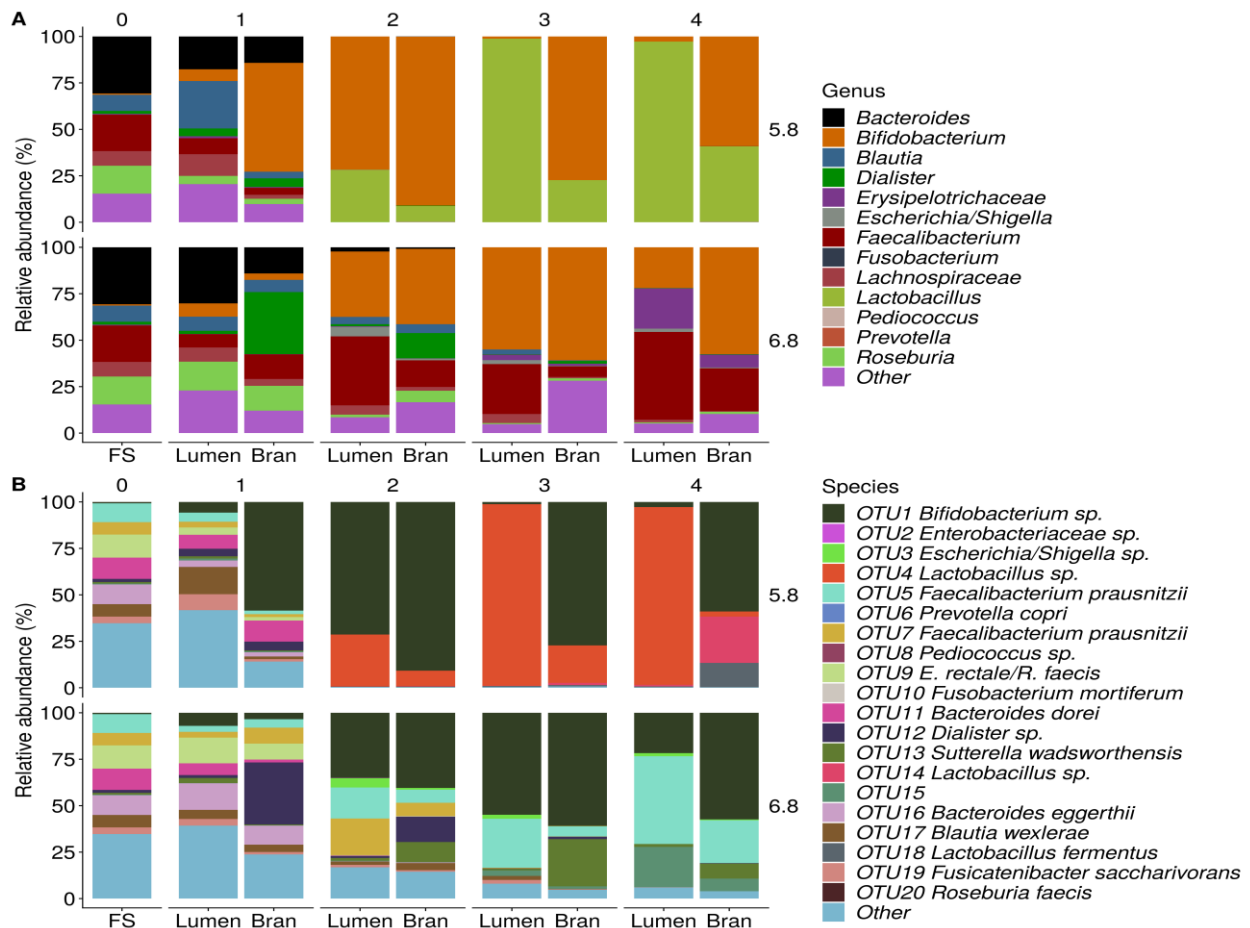



Figure 4: Shifts in genus (A) and species (B) level microbial community composition of donor 1 during consecutive enrichment steps with wheat bran as the sole nutrient source. The fecal sample (FS; enrichment step 0) was incubated with wheat bran for 24 h (enrichment step 1), after which the wheat bran residue was washed to remove loosely attached bacteria and used to seed a new incubation (enrichment step 2). This procedure was repeated two more times (enrichment step 3 and 4). Family level taxa appearing in the genus level plots should be interpreted as unclassified genus belonging to the respective family.

Genus Level

```

otuspec<- genusotutab3
otuspec <- cbind(sampleinfo3,t(otuspec))
otuspec <- subset(otuspec,Enrichment=="4"|Enrichment=="0")
otuspec <- otuspec[,-c(1:length(colnames(sampleinfo3)))]
otuspec <- data.frame(t(otuspec))
selecttop20par <- rowSums(otuspec)
otuspec <- tibble::rownames_to_column(otuspec)
otuspec <- as.data.frame(otuspec %>% top_n(13,selecttop20par))
other <- 1-colSums(otuspec[, -1])
rownames(otuspec) <- otuspec$rowname
otuspec <- otuspec[, -1]

```

```

otuspec <- rbind(otuspec,other)
rownames(otuspec)[nrow(otuspec)] <- "Other"
otuspec <- 100*otuspec

sampleinfosub3 <- subset(sampleinfo3,Enrichment=="4" | Enrichment=="0")
otuspec <- mutate(as.data.frame(t(otuspec)),donor=sampleinfosub3$Donor,LBM=sa
mpleinfosub3$Lumen_bran,enrichment=sampleinfosub3$Enrichment,pH=sampleinfosub
3$pH,names=colnames(otuspec))
otuspec <- melt(otuspec,id.vars=c('donor','LBM','enrichment','pH','names'))
otuspec23 <- subset(otuspec,donor=="2" | donor=="3")
otuspec23$donor <- mapvalues(otuspec23$donor,from=c("2","3"),to=c("Donor2","D
onor3"))

p <- ggplot(data=otuspec23,aes(x=LBM,y=as.numeric(as.character(value))))
p <- p +geom_bar(stat="identity",aes(fill=variable))
p <- papertheme(p) +facet_grid(pH~donor,drop=TRUE,scale="free_x",space="free_
x") + scale_fill_manual(values=fifteencolors,name="Genus")
p <- p+ ylab("Relative abundance (%)") + xlab(NULL) + theme(legend.text=elemen
t_text(face='italic'),strip.text.y = element_text(angle=0),legend.position="
right") +guides(fill=guide_legend(ncol=1))
bargraph23genus <- p

# Species Level
otuspec<- otutab3[,-c(1:6)]
otuspec <- cbind(sampleinfo3,t(otuspec))
otuspec <- subset(otuspec,Enrichment=="4" | Enrichment=="0")
otuspec <- otuspec[,-c(1:length(colnames(sampleinfo3)))]
otuspec <- data.frame(t(otuspec))
selecttop20par <- rowSums(otuspec)
otuspec <- tibble::rownames_to_column(otuspec)
otuspec <- as.data.frame(otuspec %>% top_n(20,selecttop20par))
other <- 1-colSums(otuspec[,-1])
rownames(otuspec) <- otuspec$rowname
otuspec <- otuspec[,-1]
otuspec <- rbind(otuspec,other)
rownames(otuspec)[nrow(otuspec)] <- "Other"
otuspec <- 100*otuspec

sampleinfosub3 <- subset(sampleinfo3,Enrichment=="4" | Enrichment=="0")
otuspec <- mutate(as.data.frame(t(otuspec)),donor=sampleinfosub3$Donor,LBM=sa
mpleinfosub3$Lumen_bran,enrichment=sampleinfosub3$Enrichment,pH=sampleinfosub
3$pH,names=colnames(otuspec))
otuspec <- melt(otuspec,id.vars=c('donor','LBM','enrichment','pH','names'))
otuspec23 <- subset(otuspec,donor=="2" | donor=="3")
otuspec23$donor <- mapvalues(otuspec23$donor,from=c("2","3"),to=c("Donor2","D
onor3"))

p <- ggplot(data=otuspec23,aes(x=LBM,y=as.numeric(as.character(value))))
p <- p +geom_bar(stat="identity",aes(fill=variable))
p <- papertheme(p) +facet_grid(pH~donor,drop=TRUE,scale="free_x",space="free_

```

```
x") + scale_fill_manual(values=thirtycolors,name="Species",labels=label3donor
sillu)
p <- p+ ylab("Relative abundance (%)") + xlab(NULL) + theme(legend.text=element_text(face='italic'),strip.text.y = element_text(angle=0),legend.position="right") +guides(fill=guide_legend(ncol=1))
bargraph23species <- p

tiff("Figure5.tiff",width=12*dpi,height=16*dpi,res=dpi,compression="lzw")
cowplot::plot_grid(bargraph23genus,bargraph23species,nrow=2,rel_heights =c(0.5,0.5),labels='AUTO',label_size=20,label_fontfamily='sans',align="v",axis="1")
dev.off()

## png
## 2

include_graphics("Figure5.tiff")
```

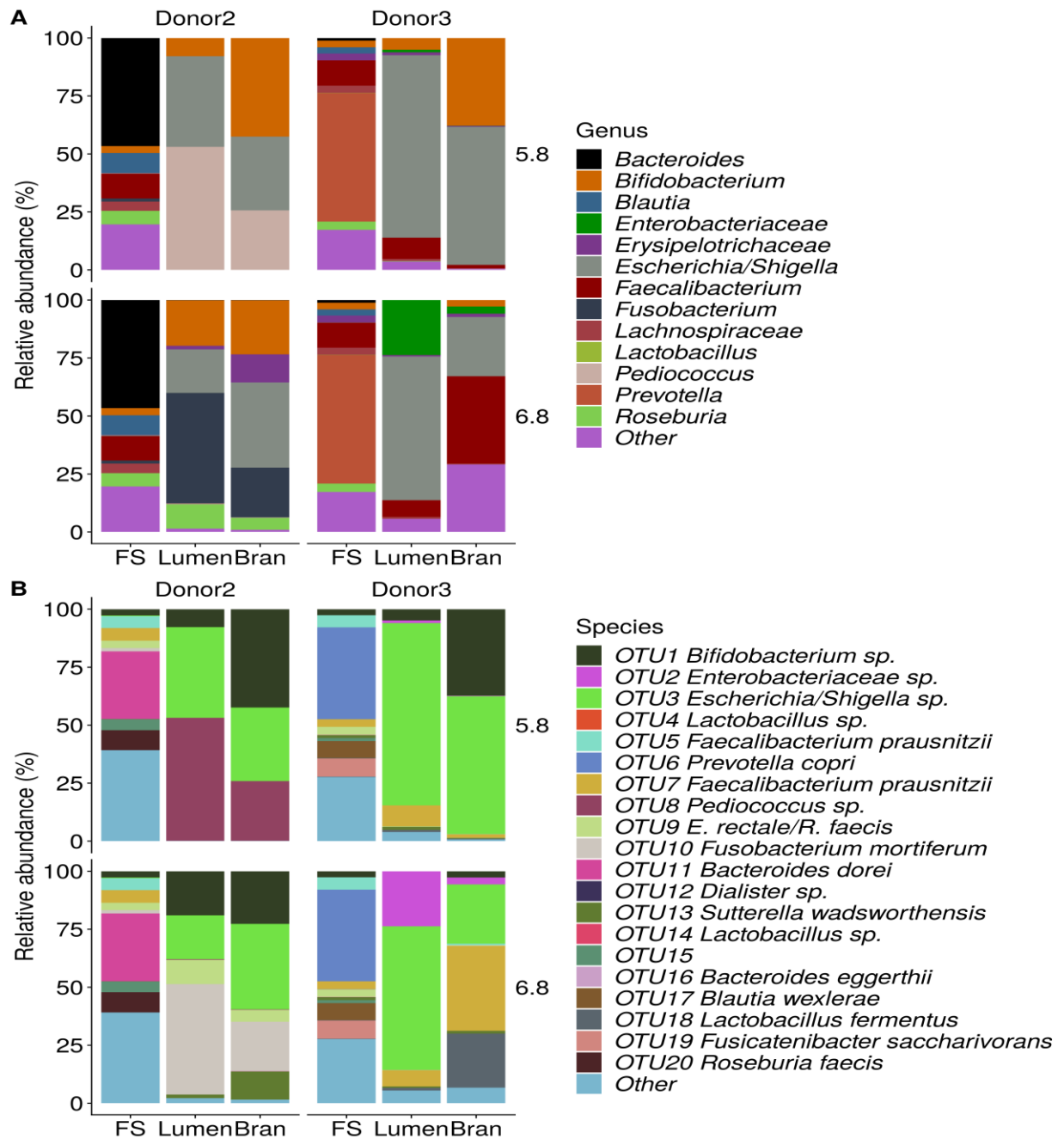


Figure 5: Shifts in genus (A) and species (B) level microbial community composition of donor 2 and 3 after four enrichment steps with wheat bran as the sole nutrient source. The fecal sample (FS; enrichment step 0) was incubated with wheat bran for 24 h (enrichment step 1), after which the wheat bran residue was washed to remove loosely attached bacteria and used to seed a new incubation (enrichment step 2). This procedure was repeated two more times (enrichment step 3 and 4). Only the final enrichment step and FS are shown in this plot. Family level taxa appearing in the genus level plots should be interpreted as unclassified genus belonging to the respective family.

```

# Define additional colors
fifteencolorsbis <- c("#C9A325", "#000000", "#CD6600", "#36648B", "#008B00", "#838B83", "#8B0000", "#323C4D", "#A03D44", "#98B736", "#7FCD50", "#BB5236", "#C8ADA4", "#AB5CC7", "#7A378B")

# Species level annotation of OTUs (verified by BLAST and RDP)
label4donorsillu <- c("OTU1 Escherichia/Shigella sp.", "OTU2 Bifidobacterium s p.", "OTU3 Faecalibacterium prausnitzii", "OTU4 Lactobacillus gasseri", "OTU5 Cl ostridium XVII", "OTU6 Lachnospiraceae sp.", "OTU7 Fusicatenibacter saccharivor ans", "OTU8 Clostridium XIVa sp.", "OTU9 Collinsella aerofaciens", "OTU10 Eubact erium rectale", "OTU11 Bacteroides vulgatus", "OTU12 Bifidobacterium longum", "O TU13 Gemmiger formicilis", "OTU14 Dialister invisus", "Other")

otuspec <- genusotutab4
selecttop20par <- rowSums(otuspec)
otuspec <- tibble::rownames_to_column(otuspec)
otuspec <- as.data.frame(otuspec %>% top_n(13, selecttop20par))
other <- 1-colSums(otuspec[, -1])
rownames(otuspec) <- otuspec$rowname
otuspec <- otuspec[, -1]
otuspec <- rbind(otuspec, other)
rownames(otuspec)[nrow(otuspec)] <- "Other"
otuspec <- otuspec*100

otuspec <- mutate(as.data.frame(t(otuspec)), donor=sampleinfo4$Donor, LBM=sampl einfo4$Lumen_bran, enrichment=sampleinfo4$Enrichment, pH=sampleinfo4$pH, names=c olnames(otuspec))
otuspec <- otuspec[c(5,6,9,10),]
otuspec4 <- melt(otuspec, id.vars=c('donor', 'LBM', 'enrichment', 'pH', 'names'))

p <- ggplot(data=otuspec4, aes(x=LBM, y=as.numeric(as.character(value))))
p <- p + geom_bar(stat="identity", aes(fill=variable))
p <- papertheme(p) + facet_grid(pH~., drop=TRUE, scale="free_x", space="free_x")
+ scale_fill_manual(values=fifteencolorsbis, name="Genus")
p <- p + ylab("Relative abundance (%)") + xlab(NULL) + theme(legend.text=elemen t_text(face='italic'), legend.position="right", strip.text.y = element_text(an gle=0)) + guides(fill=guide_legend(ncol=1))
bargraph4genus <- p

# Species
otuspec <- otutab4[, -c(1:6)]
selecttop20par <- rowSums(otuspec)
otuspec <- tibble::rownames_to_column(otuspec)
otuspec <- as.data.frame(otuspec %>% top_n(14, selecttop20par))
other <- 1-colSums(otuspec[, -1])
rownames(otuspec) <- otuspec$rowname
otuspec <- otuspec[, -1]
otuspec <- rbind(otuspec, other)
rownames(otuspec)[nrow(otuspec)] <- "Other"
otuspec <- otuspec*100

```

```

otuspec <- mutate(as.data.frame(t(otuspec)),donor=sampleinfo4$Donor,LBM=sampleinfo4$Lumen_bran,enrichment=sampleinfo4$Enrichment,pH=sampleinfo4$pH,names=colnames(otuspec))
otuspec <- otuspec[c(5,6,9,10),]
otuspec4 <- melt(otuspec,id.vars=c('donor','LBM','enrichment','pH','names'))

p <- ggplot(data=otuspec4,aes(x=LBM,y=as.numeric(as.character(value))))
p <- p +geom_bar(stat="identity",aes(fill=variable))
p <- papertheme(p) +facet_grid(pH~.,drop=TRUE,scale="free_x",space="free_x")
+ scale_fill_manual(values=thirtycolors,name="Species",label=label4donorsillum)
)
p <- p+ ylab("Relative abundance (%)") + xlab(NULL) + theme(legend.text=element_text(face='italic'),legend.position="right",strip.text.y = element_text(angle=0)) +guides(fill=guide_legend(ncol=1))
bargraph4species <- p

tiff("Figure6.tiff",width=12*dpi,height=10*dpi,res=dpi,compression="lzw")
cowplot::plot_grid(bargraph4genus,bargraph4species,nrow=2,rel_heights =c(0.5,0.5),labels='AUTO',label_size=20,label_fontfamily='sans',align="v",axis="1")
dev.off()

## png
## 2

include_graphics("Figure6.tiff")

```

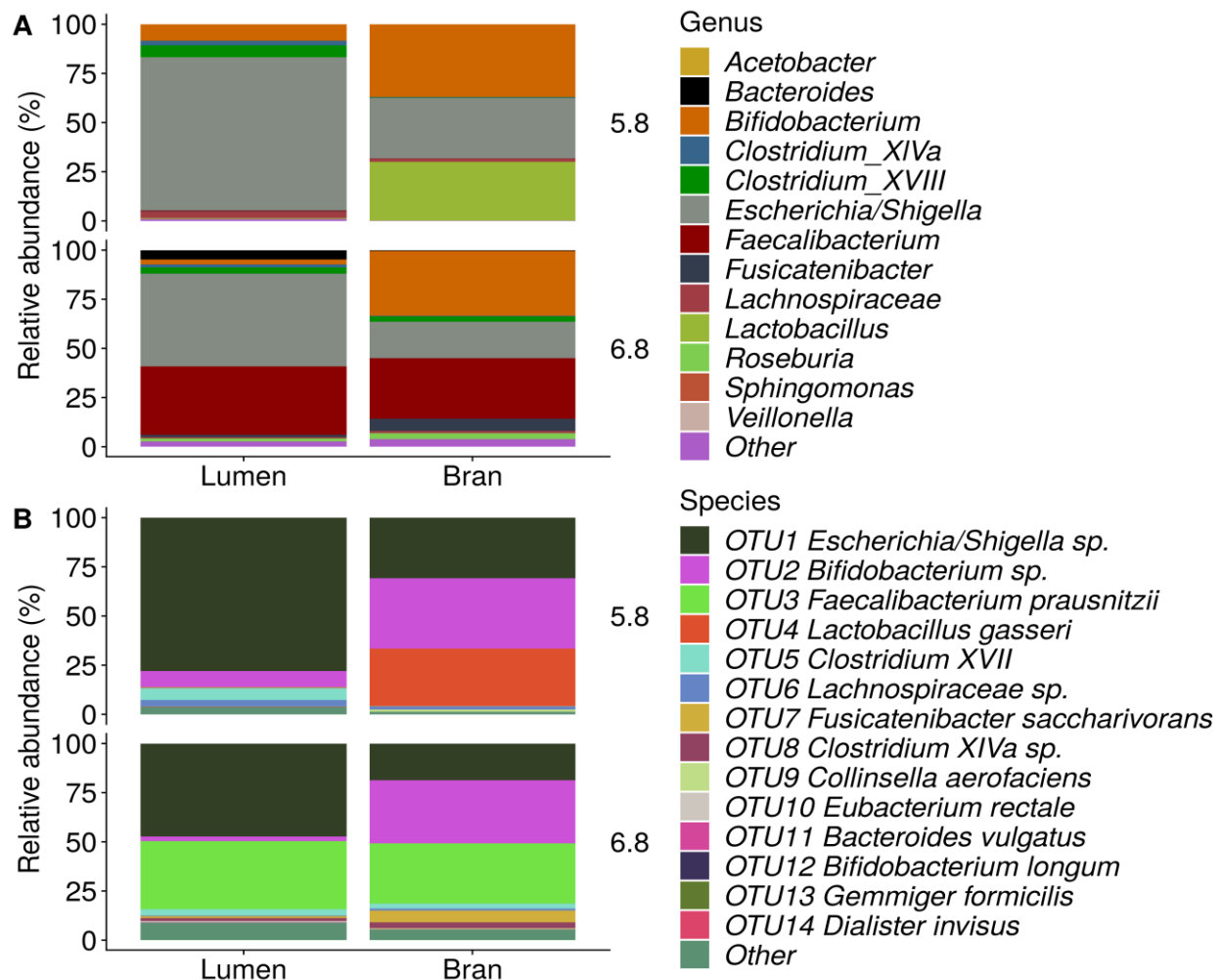


Figure 6: Genus (A) and species (B) level microbial community composition of donor 4 after the final enrichment step with wheat bran as the sole nutrient source. The fecal sample (FS; enrichment step 0) was incubated with wheat bran for 24 h (enrichment step 1), after which the wheat bran residue was washed to remove loosely attached bacteria and used to seed a new incubation (enrichment step 2). This procedure was repeated two more times (enrichment step 3 and 4). Only the final enrichment step was shown in this plot, due to a low read number (<100 reads) in the fecal sample. Family level taxa appearing in the genus level plots should be interpreted as unclassified genus belonging to the respective family.

Build consensus sanger sequences

The OTUs, resulting from 16S rRNA gene amplicon sequencing of the consecutive enrichments were compared to the 16S rRNA gene Sanger reads of the obtained isolates by means of a phylogenetic placement analysis. Forward and reverse Sanger sequences for each donor were grouped into separate files and reverse complements and summary statistics were obtained using the mothur software package (v.1.39.5) (Schloss et al. 2009). The 515F-806R primer pair, used for Illumina MiSeq 16S rRNA gene amplicon sequencing was located

in the Sanger reads (forward and reverse compliment). In case both primers were not present on one and the same read (either forward, or reverse), consensus sequences (contigs) were generated using the sangeranalyseR package (version 0.1.0) (Lanfeard 2015). Contigs with more than 100 degenerated positions, indicative of a poor quality alignment, were omitted.

```
# Summarize data
sf = summarise.abi.folder("Sanger_isolates")

## [1] "Looking for .ab1 files..."
## [1] "Found 256 .ab1 files..."
## [1] "Loading reads..."
## [1] "Calculating read summaries..."
## [1] "Cleaning up"

sf$summaries

##           file.path      folder.name  file.name  raw.length
## 1 Sanger_isolates/100F.F.ab1 Sanger_isolates 100F.F.ab1      891
## 2 Sanger_isolates/100R.R.ab1 Sanger_isolates 100R.R.ab1      485
## 3 Sanger_isolates/101F.F.ab1 Sanger_isolates 101F.F.ab1      793
## 4 Sanger_isolates/101R.R.ab1 Sanger_isolates 101R.R.ab1     1128
## 5 Sanger_isolates/102F.F.ab1 Sanger_isolates 102F.F.ab1     1164
## 6 Sanger_isolates/102R.R.ab1 Sanger_isolates 102R.R.ab1     1100
## 7 Sanger_isolates/103F.F.ab1 Sanger_isolates 103F.F.ab1     1117
## 8 Sanger_isolates/103R.R.ab1 Sanger_isolates 103R.R.ab1     1115
## 9 Sanger_isolates/104F.F.ab1 Sanger_isolates 104F.F.ab1     1165
## 10 Sanger_isolates/104R.R.ab1 Sanger_isolates 104R.R.ab1     1166
## 11 Sanger_isolates/105F.F.ab1 Sanger_isolates 105F.F.ab1     1074
## 12 Sanger_isolates/105R.R.ab1 Sanger_isolates 105R.R.ab1      805
## 13 Sanger_isolates/106F.F.ab1 Sanger_isolates 106F.F.ab1      894
## 14 Sanger_isolates/106R.R.ab1 Sanger_isolates 106R.R.ab1     1116
## 15 Sanger_isolates/107F.F.ab1 Sanger_isolates 107F.F.ab1     1021
## 16 Sanger_isolates/107R.R.ab1 Sanger_isolates 107R.R.ab1     1133
## 17 Sanger_isolates/108F.F.ab1 Sanger_isolates 108F.F.ab1      962
## 18 Sanger_isolates/108R.R.ab1 Sanger_isolates 108R.R.ab1     1088
## 19 Sanger_isolates/109F.F.ab1 Sanger_isolates 109F.F.ab1     1126
## 20 Sanger_isolates/109R.R.ab1 Sanger_isolates 109R.R.ab1     1148
## 21 Sanger_isolates/10F.F.ab1 Sanger_isolates 10F.F.ab1     1045
## 22 Sanger_isolates/10R.R.ab1 Sanger_isolates 10R.R.ab1     1104
## 23 Sanger_isolates/110F.F.ab1 Sanger_isolates 110F.F.ab1      876
## 24 Sanger_isolates/110R.R.ab1 Sanger_isolates 110R.R.ab1     1033
## 25 Sanger_isolates/11F.F.ab1 Sanger_isolates 11F.F.ab1      546
## 26 Sanger_isolates/11R.R.ab1 Sanger_isolates 11R.R.ab1      683
## 27 Sanger_isolates/128F.F.ab1 Sanger_isolates 128F.F.ab1     1017
## 28 Sanger_isolates/128R.R.ab1 Sanger_isolates 128R.R.ab1     1196
## 29 Sanger_isolates/129F.F.ab1 Sanger_isolates 129F.F.ab1      711
## 30 Sanger_isolates/129R.R.ab1 Sanger_isolates 129R.R.ab1     1073
## 31 Sanger_isolates/130F.F.ab1 Sanger_isolates 130F.F.ab1      546
## 32 Sanger_isolates/130R.R.ab1 Sanger_isolates 130R.R.ab1      489
## 33 Sanger_isolates/131F.F.ab1 Sanger_isolates 131F.F.ab1      977
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## 34	Sanger_isolates/131R.R.ab1	Sanger_isolates	131R.R.ab1	1105
## 35	Sanger_isolates/132F.F.ab1	Sanger_isolates	132F.F.ab1	200
## 36	Sanger_isolates/132R.R.ab1	Sanger_isolates	132R.R.ab1	199
## 37	Sanger_isolates/133F.F.ab1	Sanger_isolates	133F.F.ab1	199
## 38	Sanger_isolates/133R.R.ab1	Sanger_isolates	133R.R.ab1	1001
## 39	Sanger_isolates/134F.F.ab1	Sanger_isolates	134F.F.ab1	287
## 40	Sanger_isolates/134R.R.ab1	Sanger_isolates	134R.R.ab1	341
## 41	Sanger_isolates/135F.F.ab1	Sanger_isolates	135F.F.ab1	993
## 42	Sanger_isolates/135R.R.ab1	Sanger_isolates	135R.R.ab1	129
## 43	Sanger_isolates/136F.F.ab1	Sanger_isolates	136F.F.ab1	920
## 44	Sanger_isolates/136R.R.ab1	Sanger_isolates	136R.R.ab1	1109
## 45	Sanger_isolates/137F.F.ab1	Sanger_isolates	137F.F.ab1	735
## 46	Sanger_isolates/137R.R.ab1	Sanger_isolates	137R.R.ab1	691
## 47	Sanger_isolates/138F.F.ab1	Sanger_isolates	138F.F.ab1	714
## 48	Sanger_isolates/138R.R.ab1	Sanger_isolates	138R.R.ab1	872
## 49	Sanger_isolates/139F.F.ab1	Sanger_isolates	139F.F.ab1	610
## 50	Sanger_isolates/139R.R.ab1	Sanger_isolates	139R.R.ab1	382
## 51	Sanger_isolates/13F.F.ab1	Sanger_isolates	13F.F.ab1	604
## 52	Sanger_isolates/13R.R.ab1	Sanger_isolates	13R.R.ab1	957
## 53	Sanger_isolates/140F.F.ab1	Sanger_isolates	140F.F.ab1	542
## 54	Sanger_isolates/140R.R.ab1	Sanger_isolates	140R.R.ab1	943
## 55	Sanger_isolates/141F.F.ab1	Sanger_isolates	141F.F.ab1	1060
## 56	Sanger_isolates/141R.R.ab1	Sanger_isolates	141R.R.ab1	779
## 57	Sanger_isolates/142F.F.ab1	Sanger_isolates	142F.F.ab1	680
## 58	Sanger_isolates/142R.R.ab1	Sanger_isolates	142R.R.ab1	707
## 59	Sanger_isolates/143F.F.ab1	Sanger_isolates	143F.F.ab1	1067
## 60	Sanger_isolates/143R.R.ab1	Sanger_isolates	143R.R.ab1	1019
## 61	Sanger_isolates/144F.F.ab1	Sanger_isolates	144F.F.ab1	905
## 62	Sanger_isolates/144R.R.ab1	Sanger_isolates	144R.R.ab1	1059
## 63	Sanger_isolates/145F.F.ab1	Sanger_isolates	145F.F.ab1	1049
## 64	Sanger_isolates/145R.R.ab1	Sanger_isolates	145R.R.ab1	650
## 65	Sanger_isolates/146F.F.ab1	Sanger_isolates	146F.F.ab1	1121
## 66	Sanger_isolates/146R.R.ab1	Sanger_isolates	146R.R.ab1	1145
## 67	Sanger_isolates/147F.F.ab1	Sanger_isolates	147F.F.ab1	1002
## 68	Sanger_isolates/147R.R.ab1	Sanger_isolates	147R.R.ab1	1050
## 69	Sanger_isolates/148F.F.ab1	Sanger_isolates	148F.F.ab1	736
## 70	Sanger_isolates/148R.R.ab1	Sanger_isolates	148R.R.ab1	390
## 71	Sanger_isolates/149F.F.ab1	Sanger_isolates	149F.F.ab1	660
## 72	Sanger_isolates/149R.R.ab1	Sanger_isolates	149R.R.ab1	1139
## 73	Sanger_isolates/150F.F.ab1	Sanger_isolates	150F.F.ab1	505
## 74	Sanger_isolates/150R.R.ab1	Sanger_isolates	150R.R.ab1	937
## 75	Sanger_isolates/151F.F.ab1	Sanger_isolates	151F.F.ab1	706
## 76	Sanger_isolates/151R.R.ab1	Sanger_isolates	151R.R.ab1	999
## 77	Sanger_isolates/152F.F.ab1	Sanger_isolates	152F.F.ab1	614
## 78	Sanger_isolates/152R.R.ab1	Sanger_isolates	152R.R.ab1	843
## 79	Sanger_isolates/153F.F.ab1	Sanger_isolates	153F.F.ab1	703
## 80	Sanger_isolates/153R.R.ab1	Sanger_isolates	153R.R.ab1	933
## 81	Sanger_isolates/154F.F.ab1	Sanger_isolates	154F.F.ab1	943
## 82	Sanger_isolates/154R.R.ab1	Sanger_isolates	154R.R.ab1	1221
## 83	Sanger_isolates/155F.F.ab1	Sanger_isolates	155F.F.ab1	752

## 84	Sanger_isolates/155R.R.ab1	Sanger_isolates	155R.R.ab1	1122
## 85	Sanger_isolates/156F.F.ab1	Sanger_isolates	156F.F.ab1	610
## 86	Sanger_isolates/156R.R.ab1	Sanger_isolates	156R.R.ab1	1034
## 87	Sanger_isolates/157F.F.ab1	Sanger_isolates	157F.F.ab1	712
## 88	Sanger_isolates/157R.R.ab1	Sanger_isolates	157R.R.ab1	1115
## 89	Sanger_isolates/158F.F.ab1	Sanger_isolates	158F.F.ab1	874
## 90	Sanger_isolates/158R.R.ab1	Sanger_isolates	158R.R.ab1	990
## 91	Sanger_isolates/159F.F.ab1	Sanger_isolates	159F.F.ab1	702
## 92	Sanger_isolates/159R.R.ab1	Sanger_isolates	159R.R.ab1	1103
## 93	Sanger_isolates/15F.F.ab1	Sanger_isolates	15F.F.ab1	829
## 94	Sanger_isolates/15R.R.ab1	Sanger_isolates	15R.R.ab1	923
## 95	Sanger_isolates/160F.F.ab1	Sanger_isolates	160F.F.ab1	778
## 96	Sanger_isolates/160R.R.ab1	Sanger_isolates	160R.R.ab1	1157
## 97	Sanger_isolates/161F.F.ab1	Sanger_isolates	161F.F.ab1	796
## 98	Sanger_isolates/161R.R.ab1	Sanger_isolates	161R.R.ab1	1015
## 99	Sanger_isolates/162F.F.ab1	Sanger_isolates	162F.F.ab1	665
## 100	Sanger_isolates/162R.R.ab1	Sanger_isolates	162R.R.ab1	581
## 101	Sanger_isolates/163F.F.ab1	Sanger_isolates	163F.F.ab1	806
## 102	Sanger_isolates/163R.R.ab1	Sanger_isolates	163R.R.ab1	837
## 103	Sanger_isolates/181F.F.ab1	Sanger_isolates	181F.F.ab1	1149
## 104	Sanger_isolates/181R.R.ab1	Sanger_isolates	181R.R.ab1	1188
## 105	Sanger_isolates/182F.F.ab1	Sanger_isolates	182F.F.ab1	1050
## 106	Sanger_isolates/182R.R.ab1	Sanger_isolates	182R.R.ab1	815
## 107	Sanger_isolates/183F.F.ab1	Sanger_isolates	183F.F.ab1	1044
## 108	Sanger_isolates/183R.R.ab1	Sanger_isolates	183R.R.ab1	1190
## 109	Sanger_isolates/184F.F.ab1	Sanger_isolates	184F.F.ab1	916
## 110	Sanger_isolates/184R.R.ab1	Sanger_isolates	184R.R.ab1	1151
## 111	Sanger_isolates/185F.F.ab1	Sanger_isolates	185F.F.ab1	757
## 112	Sanger_isolates/185R.R.ab1	Sanger_isolates	185R.R.ab1	1147
## 113	Sanger_isolates/186F.F.ab1	Sanger_isolates	186F.F.ab1	595
## 114	Sanger_isolates/186R.R.ab1	Sanger_isolates	186R.R.ab1	1023
## 115	Sanger_isolates/187F.F.ab1	Sanger_isolates	187F.F.ab1	1142
## 116	Sanger_isolates/187R.R.ab1	Sanger_isolates	187R.R.ab1	1131
## 117	Sanger_isolates/188F.F.ab1	Sanger_isolates	188F.F.ab1	1095
## 118	Sanger_isolates/188R.R.ab1	Sanger_isolates	188R.R.ab1	1023
## 119	Sanger_isolates/189F.F.ab1	Sanger_isolates	189F.F.ab1	1049
## 120	Sanger_isolates/189R.R.ab1	Sanger_isolates	189R.R.ab1	1029
## 121	Sanger_isolates/18F.F.ab1	Sanger_isolates	18F.F.ab1	655
## 122	Sanger_isolates/18R.R.ab1	Sanger_isolates	18R.R.ab1	999
## 123	Sanger_isolates/190F.F.ab1	Sanger_isolates	190F.F.ab1	1063
## 124	Sanger_isolates/190R.R.ab1	Sanger_isolates	190R.R.ab1	709
## 125	Sanger_isolates/191F.F.ab1	Sanger_isolates	191F.F.ab1	1021
## 126	Sanger_isolates/191R.R.ab1	Sanger_isolates	191R.R.ab1	1153
## 127	Sanger_isolates/192F.F.ab1	Sanger_isolates	192F.F.ab1	1208
## 128	Sanger_isolates/192R.R.ab1	Sanger_isolates	192R.R.ab1	1157
## 129	Sanger_isolates/193F.F.ab1	Sanger_isolates	193F.F.ab1	1009
## 130	Sanger_isolates/193R.R.ab1	Sanger_isolates	193R.R.ab1	1121
## 131	Sanger_isolates/194F.F.ab1	Sanger_isolates	194F.F.ab1	1020
## 132	Sanger_isolates/194R.R.ab1	Sanger_isolates	194R.R.ab1	1137
## 133	Sanger_isolates/195F.F.ab1	Sanger_isolates	195F.F.ab1	1091

## 134	Sanger_isolates/195R.R.ab1	Sanger_isolates	195R.R.ab1	1107
## 135	Sanger_isolates/196F.F.ab1	Sanger_isolates	196F.F.ab1	1175
## 136	Sanger_isolates/196R.R.ab1	Sanger_isolates	196R.R.ab1	1095
## 137	Sanger_isolates/197F.F.ab1	Sanger_isolates	197F.F.ab1	676
## 138	Sanger_isolates/197R.R.ab1	Sanger_isolates	197R.R.ab1	1179
## 139	Sanger_isolates/198F.F.ab1	Sanger_isolates	198F.F.ab1	1100
## 140	Sanger_isolates/198R.R.ab1	Sanger_isolates	198R.R.ab1	928
## 141	Sanger_isolates/199F.F.ab1	Sanger_isolates	199F.F.ab1	609
## 142	Sanger_isolates/199R.R.ab1	Sanger_isolates	199R.R.ab1	906
## 143	Sanger_isolates/19F.F.ab1	Sanger_isolates	19F.F.ab1	1060
## 144	Sanger_isolates/19R.R.ab1	Sanger_isolates	19R.R.ab1	1160
## 145	Sanger_isolates/200F.F.ab1	Sanger_isolates	200F.F.ab1	1217
## 146	Sanger_isolates/200R.R.ab1	Sanger_isolates	200R.R.ab1	368
## 147	Sanger_isolates/201F.F.ab1	Sanger_isolates	201F.F.ab1	1183
## 148	Sanger_isolates/201R.R.ab1	Sanger_isolates	201R.R.ab1	965
## 149	Sanger_isolates/202F.F.ab1	Sanger_isolates	202F.F.ab1	1227
## 150	Sanger_isolates/202R.R.ab1	Sanger_isolates	202R.R.ab1	1117
## 151	Sanger_isolates/203F.F.ab1	Sanger_isolates	203F.F.ab1	482
## 152	Sanger_isolates/203R.R.ab1	Sanger_isolates	203R.R.ab1	987
## 153	Sanger_isolates/204F.F.ab1	Sanger_isolates	204F.F.ab1	603
## 154	Sanger_isolates/204R.R.ab1	Sanger_isolates	204R.R.ab1	584
## 155	Sanger_isolates/205F.F.ab1	Sanger_isolates	205F.F.ab1	487
## 156	Sanger_isolates/205R.R.ab1	Sanger_isolates	205R.R.ab1	566
## 157	Sanger_isolates/206F.F.ab1	Sanger_isolates	206F.F.ab1	1183
## 158	Sanger_isolates/206R.R.ab1	Sanger_isolates	206R.R.ab1	1190
## 159	Sanger_isolates/21F.F.ab1	Sanger_isolates	21F.F.ab1	903
## 160	Sanger_isolates/21R.R.ab1	Sanger_isolates	21R.R.ab1	918
## 161	Sanger_isolates/25F.F.ab1	Sanger_isolates	25F.F.ab1	904
## 162	Sanger_isolates/25R.R.ab1	Sanger_isolates	25R.R.ab1	931
## 163	Sanger_isolates/26F.F.ab1	Sanger_isolates	26F.F.ab1	1127
## 164	Sanger_isolates/26R.R.ab1	Sanger_isolates	26R.R.ab1	813
## 165	Sanger_isolates/27F.F.ab1	Sanger_isolates	27F.F.ab1	966
## 166	Sanger_isolates/27R.R.ab1	Sanger_isolates	27R.R.ab1	824
## 167	Sanger_isolates/28F.F.ab1	Sanger_isolates	28F.F.ab1	908
## 168	Sanger_isolates/28R.R.ab1	Sanger_isolates	28R.R.ab1	1017
## 169	Sanger_isolates/29F.F.ab1	Sanger_isolates	29F.F.ab1	966
## 170	Sanger_isolates/29R.R.ab1	Sanger_isolates	29R.R.ab1	1109
## 171	Sanger_isolates/30F.F.ab1	Sanger_isolates	30F.F.ab1	914
## 172	Sanger_isolates/30R.R.ab1	Sanger_isolates	30R.R.ab1	945
## 173	Sanger_isolates/31F.F.ab1	Sanger_isolates	31F.F.ab1	970
## 174	Sanger_isolates/31R.R.ab1	Sanger_isolates	31R.R.ab1	999
## 175	Sanger_isolates/32F.F.ab1	Sanger_isolates	32F.F.ab1	1045
## 176	Sanger_isolates/32R.R.ab1	Sanger_isolates	32R.R.ab1	1234
## 177	Sanger_isolates/33F.F.ab1	Sanger_isolates	33F.F.ab1	996
## 178	Sanger_isolates/33R.R.ab1	Sanger_isolates	33R.R.ab1	825
## 179	Sanger_isolates/34F.F.ab1	Sanger_isolates	34F.F.ab1	933
## 180	Sanger_isolates/34R.R.ab1	Sanger_isolates	34R.R.ab1	1143
## 181	Sanger_isolates/35F.F.ab1	Sanger_isolates	35F.F.ab1	920
## 182	Sanger_isolates/35R.R.ab1	Sanger_isolates	35R.R.ab1	1161
## 183	Sanger_isolates/36F.F.ab1	Sanger_isolates	36F.F.ab1	843

## 184	Sanger_isolates/36R.R.ab1	Sanger_isolates	36R.R.ab1	1091
## 185	Sanger_isolates/37F.F.ab1	Sanger_isolates	37F.F.ab1	942
## 186	Sanger_isolates/37R.R.ab1	Sanger_isolates	37R.R.ab1	1018
## 187	Sanger_isolates/38F.F.ab1	Sanger_isolates	38F.F.ab1	771
## 188	Sanger_isolates/38R.R.ab1	Sanger_isolates	38R.R.ab1	610
## 189	Sanger_isolates/39F.F.ab1	Sanger_isolates	39F.F.ab1	481
## 190	Sanger_isolates/39R.R.ab1	Sanger_isolates	39R.R.ab1	460
## 191	Sanger_isolates/40F.F.ab1	Sanger_isolates	40F.F.ab1	1039
## 192	Sanger_isolates/40R.R.ab1	Sanger_isolates	40R.R.ab1	1046
## 193	Sanger_isolates/41F.F.ab1	Sanger_isolates	41F.F.ab1	1172
## 194	Sanger_isolates/41R.R.ab1	Sanger_isolates	41R.R.ab1	1114
## 195	Sanger_isolates/42F.F.ab1	Sanger_isolates	42F.F.ab1	988
## 196	Sanger_isolates/42R.R.ab1	Sanger_isolates	42R.R.ab1	980
## 197	Sanger_isolates/43F.F.ab1	Sanger_isolates	43F.F.ab1	1119
## 198	Sanger_isolates/43R.R.ab1	Sanger_isolates	43R.R.ab1	1164
## 199	Sanger_isolates/44F.F.ab1	Sanger_isolates	44F.F.ab1	956
## 200	Sanger_isolates/44R.R.ab1	Sanger_isolates	44R.R.ab1	1067
## 201	Sanger_isolates/45F.F.ab1	Sanger_isolates	45F.F.ab1	1088
## 202	Sanger_isolates/45R.R.ab1	Sanger_isolates	45R.R.ab1	878
## 203	Sanger_isolates/46F.F.ab1	Sanger_isolates	46F.F.ab1	990
## 204	Sanger_isolates/46R.R.ab1	Sanger_isolates	46R.R.ab1	1086
## 205	Sanger_isolates/47F.F.ab1	Sanger_isolates	47F.F.ab1	752
## 206	Sanger_isolates/47R.R.ab1	Sanger_isolates	47R.R.ab1	582
## 207	Sanger_isolates/48F.F.ab1	Sanger_isolates	48F.F.ab1	1157
## 208	Sanger_isolates/48R.R.ab1	Sanger_isolates	48R.R.ab1	200
## 209	Sanger_isolates/76F.F.ab1	Sanger_isolates	76F.F.ab1	1138
## 210	Sanger_isolates/76R.R.ab1	Sanger_isolates	76R.R.ab1	1168
## 211	Sanger_isolates/77F.F.ab1	Sanger_isolates	77F.F.ab1	1095
## 212	Sanger_isolates/77R.R.ab1	Sanger_isolates	77R.R.ab1	1111
## 213	Sanger_isolates/78F.F.ab1	Sanger_isolates	78F.F.ab1	944
## 214	Sanger_isolates/78R.R.ab1	Sanger_isolates	78R.R.ab1	1180
## 215	Sanger_isolates/79F.F.ab1	Sanger_isolates	79F.F.ab1	528
## 216	Sanger_isolates/79R.R.ab1	Sanger_isolates	79R.R.ab1	966
## 217	Sanger_isolates/80F.F.ab1	Sanger_isolates	80F.F.ab1	638
## 218	Sanger_isolates/80R.R.ab1	Sanger_isolates	80R.R.ab1	999
## 219	Sanger_isolates/81F.F.ab1	Sanger_isolates	81F.F.ab1	801
## 220	Sanger_isolates/81R.R.ab1	Sanger_isolates	81R.R.ab1	1031
## 221	Sanger_isolates/82F.F.ab1	Sanger_isolates	82F.F.ab1	891
## 222	Sanger_isolates/82R.R.ab1	Sanger_isolates	82R.R.ab1	1129
## 223	Sanger_isolates/83F.F.ab1	Sanger_isolates	83F.F.ab1	637
## 224	Sanger_isolates/83R.R.ab1	Sanger_isolates	83R.R.ab1	915
## 225	Sanger_isolates/84F.F.ab1	Sanger_isolates	84F.F.ab1	515
## 226	Sanger_isolates/84R.R.ab1	Sanger_isolates	84R.R.ab1	1088
## 227	Sanger_isolates/85F.F.ab1	Sanger_isolates	85F.F.ab1	525
## 228	Sanger_isolates/85R.R.ab1	Sanger_isolates	85R.R.ab1	1072
## 229	Sanger_isolates/86F.F.ab1	Sanger_isolates	86F.F.ab1	513
## 230	Sanger_isolates/86R.R.ab1	Sanger_isolates	86R.R.ab1	1104
## 231	Sanger_isolates/87F.F.ab1	Sanger_isolates	87F.F.ab1	699
## 232	Sanger_isolates/87R.R.ab1	Sanger_isolates	87R.R.ab1	1022
## 233	Sanger_isolates/88F.F.ab1	Sanger_isolates	88F.F.ab1	990

##	234	Sanger_isolates/88R.R.ab1	Sanger_isolates	88R.R.ab1	820
##	235	Sanger_isolates/89F.F.ab1	Sanger_isolates	89F.F.ab1	628
##	236	Sanger_isolates/89R.R.ab1	Sanger_isolates	89R.R.ab1	916
##	237	Sanger_isolates/90F.F.ab1	Sanger_isolates	90F.F.ab1	950
##	238	Sanger_isolates/90R.R.ab1	Sanger_isolates	90R.R.ab1	1182
##	239	Sanger_isolates/91F.F.ab1	Sanger_isolates	91F.F.ab1	973
##	240	Sanger_isolates/91R.R.ab1	Sanger_isolates	91R.R.ab1	1112
##	241	Sanger_isolates/92F.F.ab1	Sanger_isolates	92F.F.ab1	988
##	242	Sanger_isolates/92R.R.ab1	Sanger_isolates	92R.R.ab1	1127
##	243	Sanger_isolates/93F.F.ab1	Sanger_isolates	93F.F.ab1	933
##	244	Sanger_isolates/93R.R.ab1	Sanger_isolates	93R.R.ab1	1049
##	245	Sanger_isolates/94F.F.ab1	Sanger_isolates	94F.F.ab1	986
##	246	Sanger_isolates/94R.R.ab1	Sanger_isolates	94R.R.ab1	802
##	247	Sanger_isolates/95F.F.ab1	Sanger_isolates	95F.F.ab1	1050
##	248	Sanger_isolates/95R.R.ab1	Sanger_isolates	95R.R.ab1	911
##	249	Sanger_isolates/96F.F.ab1	Sanger_isolates	96F.F.ab1	1167
##	250	Sanger_isolates/96R.R.ab1	Sanger_isolates	96R.R.ab1	1125
##	251	Sanger_isolates/97F.F.ab1	Sanger_isolates	97F.F.ab1	915
##	252	Sanger_isolates/97R.R.ab1	Sanger_isolates	97R.R.ab1	993
##	253	Sanger_isolates/98F.F.ab1	Sanger_isolates	98F.F.ab1	713
##	254	Sanger_isolates/98R.R.ab1	Sanger_isolates	98R.R.ab1	716
##	255	Sanger_isolates/99F.F.ab1	Sanger_isolates	99F.F.ab1	1178
##	256	Sanger_isolates/99R.R.ab1	Sanger_isolates	99R.R.ab1	1116
##		trimmed.length	trim.start	trim.finish	raw.secondary.peaks
##	1	314	24	337	264
##	2	2	484	485	237
##	3	610	20	629	27
##	4	750	21	770	8
##	5	573	20	592	38
##	6	474	73	546	20
##	7	625	14	638	25
##	8	492	14	505	12
##	9	557	17	573	32
##	10	569	26	594	29
##	11	624	14	637	39
##	12	327	5	331	74
##	13	589	14	602	16
##	14	499	6	504	7
##	15	624	11	634	33
##	16	498	5	502	32
##	17	609	12	620	23
##	18	493	13	505	26
##	19	604	10	613	20
##	20	490	15	504	42
##	21	610	11	620	55
##	22	752	16	767	24
##	23	621	11	631	8
##	24	491	15	505	19
##	25	319	143	461	209
##	26	546	39	584	238

## 27	363	11	373	58
## 28	730	17	746	38
## 29	356	17	372	55
## 30	730	19	748	29
## 31	344	23	366	26
## 32	310	14	323	16
## 33	408	85	492	265
## 34	825	19	843	23
## 35	0	0	0	145
## 36	0	0	0	136
## 37	0	0	0	147
## 38	754	16	769	25
## 39	0	0	0	223
## 40	1	341	341	243
## 41	168	22	189	177
## 42	0	0	0	99
## 43	214	16	229	201
## 44	832	21	852	14
## 45	566	17	582	19
## 46	594	15	608	24
## 47	237	192	428	204
## 48	463	18	480	35
## 49	231	34	264	56
## 50	122	17	138	23
## 51	355	120	474	81
## 52	462	40	501	86
## 53	343	37	379	31
## 54	457	54	510	150
## 55	365	11	375	55
## 56	204	17	220	38
## 57	498	25	522	40
## 58	307	14	320	42
## 59	560	20	579	116
## 60	829	18	846	39
## 61	594	23	616	47
## 62	729	19	747	28
## 63	354	22	375	57
## 64	204	18	221	43
## 65	72	83	154	174
## 66	797	19	815	16
## 67	353	22	374	54
## 68	730	15	744	22
## 69	225	277	501	264
## 70	38	10	47	232
## 71	559	23	581	17
## 72	707	18	724	26
## 73	423	17	439	6
## 74	753	17	769	23
## 75	256	13	268	271
## 76	770	19	788	14

## 77	294	85	378	104
## 78	606	18	623	16
## 79	407	78	484	121
## 80	719	14	732	22
## 81	408	85	492	156
## 82	718	19	736	29
## 83	526	12	537	38
## 84	753	16	768	22
## 85	295	85	379	88
## 86	737	20	756	8
## 87	295	85	379	154
## 88	606	19	624	7
## 89	68	87	154	161
## 90	730	18	747	40
## 91	295	81	375	124
## 92	715	19	733	26
## 93	404	86	489	131
## 94	482	30	511	36
## 95	351	26	376	149
## 96	732	18	749	23
## 97	68	86	153	160
## 98	617	18	634	5
## 99	352	27	378	112
## 100	299	12	310	135
## 101	111	20	130	223
## 102	612	68	679	32
## 103	759	7	765	17
## 104	727	18	744	33
## 105	706	14	719	28
## 106	536	11	546	25
## 107	287	240	526	242
## 108	847	20	866	111
## 109	403	21	423	286
## 110	589	17	605	21
## 111	110	17	126	313
## 112	588	17	604	19
## 113	100	31	130	105
## 114	589	15	603	11
## 115	713	18	730	11
## 116	830	16	845	10
## 117	695	17	711	18
## 118	839	14	852	9
## 119	679	16	694	32
## 120	818	22	839	20
## 121	48	74	121	115
## 122	506	6	511	24
## 123	712	15	726	34
## 124	539	15	553	33
## 125	782	17	798	22
## 126	840	10	849	14

## 127	712	18	729	41
## 128	890	11	900	15
## 129	678	21	698	16
## 130	788	8	795	43
## 131	623	12	634	49
## 132	499	8	506	9
## 133	534	18	551	12
## 134	693	14	706	13
## 135	708	18	725	34
## 136	689	18	706	7
## 137	345	325	669	461
## 138	830	15	844	22
## 139	703	6	708	47
## 140	319	10	328	267
## 141	4	388	391	438
## 142	690	17	706	28
## 143	629	11	639	77
## 144	516	15	530	25
## 145	592	18	609	31
## 146	1	368	368	243
## 147	713	8	720	9
## 148	687	20	706	32
## 149	705	15	719	22
## 150	922	18	939	13
## 151	0	0	0	331
## 152	440	13	452	118
## 153	1	603	603	431
## 154	28	14	41	361
## 155	0	0	0	380
## 156	0	0	0	416
## 157	718	15	732	13
## 158	963	17	979	21
## 159	75	20	94	411
## 160	465	11	475	48
## 161	561	74	634	107
## 162	748	19	766	30
## 163	697	14	710	24
## 164	610	13	622	5
## 165	539	39	577	48
## 166	349	15	363	31
## 167	245	34	278	262
## 168	619	17	635	14
## 169	588	7	594	44
## 170	738	1	738	24
## 171	615	14	628	83
## 172	716	11	726	14
## 173	621	19	639	25
## 174	582	21	602	30
## 175	642	25	666	24
## 176	831	9	839	33

## 177	586	11	596	24
## 178	616	9	624	7
## 179	360	99	458	126
## 180	761	23	783	35
## 181	560	21	580	29
## 182	763	10	772	19
## 183	512	16	527	33
## 184	643	19	661	24
## 185	633	21	653	25
## 186	727	16	742	4
## 187	519	10	528	63
## 188	411	17	427	20
## 189	342	15	356	31
## 190	72	15	86	207
## 191	645	10	654	29
## 192	783	18	800	11
## 193	713	14	726	71
## 194	747	17	763	12
## 195	629	11	639	44
## 196	586	14	599	54
## 197	641	15	655	17
## 198	828	14	841	22
## 199	499	23	521	56
## 200	740	15	754	14
## 201	585	16	600	39
## 202	317	34	350	199
## 203	550	17	566	31
## 204	700	30	729	29
## 205	547	20	566	28
## 206	314	13	326	33
## 207	524	14	537	57
## 208	0	0	0	129
## 209	706	9	714	11
## 210	839	14	852	14
## 211	215	46	260	367
## 212	636	33	668	268
## 213	633	14	646	30
## 214	716	28	743	12
## 215	225	15	239	107
## 216	751	23	773	12
## 217	441	76	516	114
## 218	275	26	300	47
## 219	567	7	573	64
## 220	796	21	816	17
## 221	556	10	565	28
## 222	710	17	726	15
## 223	594	21	614	12
## 224	709	33	741	25
## 225	501	14	514	0
## 226	769	24	792	30

## 227	291	10	300	27
## 228	729	17	745	12
## 229	277	22	298	4
## 230	819	21	839	26
## 231	474	25	498	41
## 232	713	17	729	9
## 233	617	8	624	67
## 234	655	19	673	42
## 235	40	139	178	220
## 236	30	30	59	340
## 237	283	58	340	105
## 238	901	19	919	13
## 239	709	9	717	24
## 240	654	18	671	16
## 241	614	16	629	13
## 242	911	10	920	10
## 243	705	17	721	12
## 244	706	15	720	10
## 245	199	76	274	280
## 246	581	23	603	33
## 247	81	70	150	205
## 248	198	29	226	33
## 249	610	14	623	46
## 250	723	20	742	13
## 251	476	217	692	126
## 252	85	55	139	204
## 253	534	15	548	38
## 254	595	19	613	3
## 255	680	17	696	28
## 256	909	17	925	14
##	trimmed.secondary.peaks	raw.mean.quality	trimmed.mean.quality	
## 1		68	23.52626	28.95860
## 2		1	17.29835	41.00000
## 3		2	45.56604	51.27049
## 4		0	47.80229	52.50400
## 5		5	38.44748	45.30890
## 6		1	41.82790	50.24684
## 7		0	44.18926	52.12320
## 8		1	46.71632	52.29472
## 9		6	42.88615	50.83124
## 10		1	42.21843	48.74868
## 11		0	43.04356	51.55609
## 12		8	44.62283	53.20489
## 13		0	45.51059	51.64686
## 14		3	46.27200	51.16836
## 15		0	42.58244	51.13141
## 16		4	43.72024	51.44309
## 17		0	44.96687	51.40558
## 18		1	45.29670	52.06288
## 19		0	43.70712	51.82616

## 20	3	44.36121	51.89796
## 21	1	41.73789	51.63607
## 22	0	46.10872	52.54122
## 23	1	46.69818	51.25282
## 24	4	44.35714	49.69654
## 25	122	19.52190	20.86834
## 26	202	21.88141	22.33516
## 27	2	41.69990	50.81267
## 28	4	45.30661	51.90548
## 29	2	44.29032	50.97472
## 30	5	44.81320	51.05342
## 31	1	42.53370	48.79651
## 32	0	47.19084	57.95161
## 33	65	25.31256	33.45588
## 34	0	46.69182	52.10182
## 35	0	11.50000	NA
## 36	0	9.68500	NA
## 37	0	9.54000	NA
## 38	0	46.45817	51.85942
## 39	0	13.90941	NA
## 40	0	16.65982	51.00000
## 41	0	33.66131	48.80952
## 42	0	13.94574	NA
## 43	29	26.37595	35.11682
## 44	0	47.07706	52.07332
## 45	1	50.60893	56.80212
## 46	0	47.13833	50.88047
## 47	41	26.01257	33.25738
## 48	11	44.81257	49.79482
## 49	20	40.38725	44.09957
## 50	5	40.43455	39.86885
## 51	21	36.43493	43.17465
## 52	12	39.42961	47.54762
## 53	6	39.91544	44.66181
## 54	69	35.06421	39.51204
## 55	0	40.40994	51.49041
## 56	1	43.90269	50.26961
## 57	2	42.61054	48.40161
## 58	0	41.78200	50.64169
## 59	0	37.81471	51.78393
## 60	9	45.50294	50.45838
## 61	1	40.44714	48.74242
## 62	3	44.63526	51.66255
## 63	1	42.36053	50.97740
## 64	0	42.45929	50.63725
## 65	11	28.17019	35.50000
## 66	0	46.61232	51.95107
## 67	3	42.39801	50.27479
## 68	5	45.96314	51.26849
## 69	63	19.39213	23.85778

## 70	9	14.86667	23.94737
## 71	1	45.83409	49.19857
## 72	3	43.60435	51.69731
## 73	1	45.42436	48.10875
## 74	1	47.12354	51.25365
## 75	84	26.38418	27.64062
## 76	1	47.26456	52.04545
## 77	47	30.14821	32.12585
## 78	0	47.72393	51.65842
## 79	64	30.43121	33.80344
## 80	0	46.96162	51.80807
## 81	66	29.17797	33.41176
## 82	0	44.49638	53.04735
## 83	1	43.10359	48.40684
## 84	3	44.52515	51.00398
## 85	41	31.71031	34.21695
## 86	0	47.69884	52.05020
## 87	49	29.23843	33.68136
## 88	0	46.78559	52.96205
## 89	11	29.88686	34.75000
## 90	0	45.77387	52.00548
## 91	44	30.51278	33.48136
## 92	0	45.26036	52.20839
## 93	10	39.76145	53.92079
## 94	7	45.59307	50.04357
## 95	61	30.09487	32.08547
## 96	0	44.83675	52.36885
## 97	14	29.22585	34.32353
## 98	0	47.66243	52.77958
## 99	54	31.28186	31.79830
## 100	3	38.86575	52.85284
## 101	3	26.53713	47.02703
## 102	9	43.79380	48.55229
## 103	0	45.57021	51.87352
## 104	1	45.90696	52.61073
## 105	2	46.25922	51.52691
## 106	2	49.07506	57.05784
## 107	64	26.86437	30.41812
## 108	22	39.32750	48.58796
## 109	93	23.98364	30.43921
## 110	1	46.42845	52.27844
## 111	31	26.95376	32.40909
## 112	1	45.30882	51.76531
## 113	19	31.10756	33.68000
## 114	3	48.83965	52.37012
## 115	0	46.89791	52.07293
## 116	2	46.71604	51.49157
## 117	7	45.79581	51.01007
## 118	5	49.07415	50.99046
## 119	4	44.54502	50.10457

## 120	2	46.13967	50.09169
## 121	3	37.86280	50.04167
## 122	12	44.37313	48.00988
## 123	3	44.11610	50.15730
## 124	3	49.96474	56.99629
## 125	3	47.09756	51.26854
## 126	1	46.81638	51.57976
## 127	0	44.17473	52.06882
## 128	2	46.29256	51.57191
## 129	0	45.55676	51.15782
## 130	1	43.72099	50.80584
## 131	0	42.37877	51.09952
## 132	0	45.78534	51.83768
## 133	0	45.72338	51.79213
## 134	0	47.28150	52.48052
## 135	0	44.07912	51.60311
## 136	0	48.79946	53.03048
## 137	243	13.87130	13.66087
## 138	1	45.91261	52.04699
## 139	0	43.39892	50.79232
## 140	100	27.14393	29.44201
## 141	4	13.86864	32.25000
## 142	1	46.50494	51.06957
## 143	53	34.43218	37.30366
## 144	4	42.84799	51.32558
## 145	2	44.51629	52.33277
## 146	1	14.18699	45.00000
## 147	2	46.45987	51.45302
## 148	3	46.46591	51.60408
## 149	1	45.02753	51.66099
## 150	3	47.18482	51.46095
## 151	0	13.25934	NA
## 152	23	38.89889	48.61364
## 153	0	14.32338	42.00000
## 154	0	15.65925	46.64286
## 155	0	12.86242	NA
## 156	0	12.37633	NA
## 157	1	45.42989	52.26741
## 158	2	46.64190	52.01454
## 159	2	17.59757	39.37333
## 160	20	43.77935	46.73763
## 161	28	33.30795	39.86631
## 162	6	45.12286	49.24465
## 163	0	43.64877	52.24821
## 164	0	51.26779	57.74098
## 165	5	41.28306	48.91651
## 166	5	47.21540	51.14900
## 167	77	26.96061	30.80408
## 168	3	45.81703	50.77544
## 169	0	42.57407	52.03741

## 170	0	44.61325	52.06967
## 171	3	40.12092	50.23902
## 172	3	47.72076	51.59916
## 173	1	44.13673	52.07729
## 174	2	45.81655	51.86770
## 175	1	45.08815	52.33645
## 176	1	43.88818	52.10469
## 177	0	44.15400	51.71843
## 178	0	47.25962	50.52922
## 179	51	32.44124	35.23889
## 180	6	44.07605	50.28252
## 181	1	45.26030	51.95179
## 182	0	45.71769	52.24771
## 183	0	41.94208	48.58203
## 184	1	43.80748	49.44946
## 185	0	45.64942	52.08057
## 186	0	47.88824	52.20083
## 187	3	45.16516	52.90366
## 188	4	45.52700	50.73966
## 189	0	42.61411	48.48538
## 190	0	23.30952	45.31944
## 191	0	46.01811	52.24031
## 192	0	47.42912	52.64112
## 193	1	40.15228	50.86816
## 194	0	47.17663	52.30254
## 195	2	42.96177	49.65978
## 196	2	45.09729	55.10922
## 197	1	45.53771	52.27769
## 198	2	45.97950	52.40580
## 199	2	43.78778	54.23246
## 200	0	46.84308	51.78784
## 201	0	43.03297	51.89744
## 202	30	25.50342	29.71609
## 203	15	41.75907	44.21818
## 204	9	41.05775	46.70000
## 205	1	43.88977	49.17733
## 206	1	42.77949	51.51592
## 207	0	39.53012	49.66985
## 208	0	9.80500	NA
## 209	0	44.99567	52.06516
## 210	2	43.39237	50.66031
## 211	68	20.19143	23.00465
## 212	195	25.31847	25.66195
## 213	4	48.50936	55.63507
## 214	1	46.05228	51.41480
## 215	2	40.14205	47.48889
## 216	2	48.06082	51.01731
## 217	72	32.29844	35.15873
## 218	4	41.25222	47.86909
## 219	1	48.68897	58.84656

## 220	2	48.70281	52.08794
## 221	3	49.28101	57.92086
## 222	0	48.26678	53.06479
## 223	5	45.04702	46.95286
## 224	11	45.16576	47.93794
## 225	0	51.21470	52.21557
## 226	4	44.86447	49.32900
## 227	2	45.89184	51.19931
## 228	2	48.61860	52.56927
## 229	1	48.41942	51.59567
## 230	1	47.16937	52.70574
## 231	0	42.68714	51.18987
## 232	0	47.34732	49.84853
## 233	13	42.83585	50.72123
## 234	0	45.38929	51.47939
## 235	4	23.82194	38.97500
## 236	0	20.64130	50.10000
## 237	26	34.88655	41.56184
## 238	1	46.73662	51.73030
## 239	0	46.37755	51.01693
## 240	0	47.77907	53.06269
## 241	0	48.38153	52.37296
## 242	0	48.85915	52.58836
## 243	0	47.27079	50.78440
## 244	3	48.34414	51.65864
## 245	71	28.73834	34.01005
## 246	13	42.83602	45.78657
## 247	2	29.74430	45.71605
## 248	3	41.85137	46.61111
## 249	2	40.89957	49.95738
## 250	4	46.03530	51.77593
## 251	54	33.62568	37.63445
## 252	14	28.39196	32.38824
## 253	1	43.49930	51.18539
## 254	0	50.22981	52.82689
## 255	4	44.20185	51.02059
## 256	2	46.10993	51.00880
##	raw.min.quality	trimmed.min.quality	
## 1	2	8	
## 2	2	41	
## 3	2	15	
## 4	2	10	
## 5	2	2	
## 6	2	23	
## 7	2	7	
## 8	2	7	
## 9	2	7	
## 10	2	2	
## 11	2	7	
## 12	2	7	

## 13	2	7
## 14	2	9
## 15	2	7
## 16	2	10
## 17	2	7
## 18	2	15
## 19	2	7
## 20	2	10
## 21	2	10
## 22	2	2
## 23	7	7
## 24	2	6
## 25	2	7
## 26	2	2
## 27	4	12
## 28	3	3
## 29	2	9
## 30	4	8
## 31	4	16
## 32	5	9
## 33	2	10
## 34	3	9
## 35	4	NA
## 36	3	NA
## 37	3	NA
## 38	5	14
## 39	5	NA
## 40	6	51
## 41	5	20
## 42	5	NA
## 43	2	7
## 44	2	22
## 45	5	5
## 46	7	22
## 47	2	8
## 48	5	7
## 49	5	5
## 50	5	8
## 51	2	7
## 52	2	6
## 53	4	12
## 54	4	4
## 55	2	12
## 56	3	6
## 57	5	15
## 58	5	7
## 59	5	20
## 60	3	10
## 61	4	15
## 62	5	14

## 63	5	20
## 64	5	6
## 65	2	12
## 66	5	9
## 67	4	9
## 68	5	8
## 69	2	5
## 70	5	9
## 71	4	14
## 72	5	14
## 73	5	8
## 74	5	14
## 75	4	8
## 76	5	9
## 77	2	8
## 78	5	9
## 79	5	10
## 80	3	9
## 81	2	10
## 82	2	9
## 83	3	20
## 84	2	14
## 85	2	10
## 86	4	8
## 87	5	12
## 88	4	8
## 89	6	12
## 90	4	8
## 91	2	10
## 92	4	7
## 93	2	31
## 94	2	7
## 95	2	7
## 96	2	8
## 97	6	12
## 98	2	14
## 99	3	7
## 100	5	9
## 101	6	14
## 102	5	12
## 103	5	8
## 104	5	12
## 105	5	5
## 106	5	5
## 107	2	8
## 108	5	8
## 109	2	7
## 110	5	10
## 111	5	10
## 112	2	14

## 113	5	10
## 114	5	8
## 115	4	11
## 116	2	2
## 117	5	5
## 118	4	4
## 119	3	5
## 120	5	5
## 121	2	24
## 122	2	8
## 123	2	4
## 124	5	8
## 125	5	6
## 126	5	9
## 127	5	20
## 128	2	18
## 129	2	2
## 130	5	5
## 131	5	20
## 132	2	5
## 133	6	11
## 134	4	9
## 135	5	14
## 136	5	9
## 137	5	5
## 138	3	10
## 139	5	5
## 140	4	10
## 141	5	19
## 142	2	12
## 143	2	7
## 144	2	7
## 145	6	20
## 146	5	45
## 147	6	8
## 148	5	5
## 149	4	9
## 150	4	4
## 151	5	NA
## 152	5	8
## 153	5	42
## 154	5	15
## 155	5	NA
## 156	5	NA
## 157	5	8
## 158	5	8
## 159	2	8
## 160	2	6
## 161	2	10
## 162	2	2

## 163	2	20
## 164	2	12
## 165	2	7
## 166	2	2
## 167	2	8
## 168	2	6
## 169	2	7
## 170	2	10
## 171	2	12
## 172	2	8
## 173	2	20
## 174	2	8
## 175	2	10
## 176	2	2
## 177	2	15
## 178	2	2
## 179	2	10
## 180	2	7
## 181	2	20
## 182	2	2
## 183	2	14
## 184	2	6
## 185	2	20
## 186	2	2
## 187	2	9
## 188	2	2
## 189	2	7
## 190	5	10
## 191	2	7
## 192	2	14
## 193	2	20
## 194	2	20
## 195	2	6
## 196	2	2
## 197	2	14
## 198	2	8
## 199	2	6
## 200	2	6
## 201	2	14
## 202	2	2
## 203	2	8
## 204	2	9
## 205	6	15
## 206	6	6
## 207	2	20
## 208	3	NA
## 209	2	9
## 210	2	2
## 211	2	7
## 212	2	8

## 213	2	2
## 214	2	7
## 215	2	2
## 216	2	2
## 217	6	8
## 218	2	7
## 219	2	13
## 220	2	10
## 221	2	14
## 222	2	12
## 223	6	7
## 224	2	14
## 225	2	15
## 226	2	8
## 227	2	14
## 228	2	12
## 229	7	12
## 230	2	9
## 231	2	20
## 232	2	15
## 233	2	7
## 234	2	22
## 235	2	14
## 236	2	40
## 237	2	10
## 238	2	2
## 239	2	14
## 240	2	7
## 241	2	15
## 242	2	13
## 243	2	14
## 244	2	7
## 245	2	13
## 246	6	10
## 247	2	21
## 248	2	2
## 249	2	2
## 250	2	6
## 251	2	10
## 252	2	2
## 253	2	15
## 254	2	8
## 255	2	7
## 256	2	2

```

p<- ggplot(sf$summaries, aes(x = folder.name, y = raw.mean.quality)) + geom_boxplot()
p <- ggplot(sf$summaries, aes(x = folder.name, y = trimmed.mean.quality)) + geom_boxplot()
p <- ggplot(sf$summaries, aes(x = folder.name, y = trimmed.secondary.peaks))

```

```

+ geom_boxplot() + geom_hline(yintercept = 2, linetype = 3, colour = 'red')
p <- ggplot(sf$summaries, aes(x = trimmed.mean.quality, y = trimmed.secondary
.peaks)) + geom_point()

# Make consensus by merging reads
fwd = readsangerseq("Sanger_isolates/13F.F.ab1")
rev = readsangerseq("Sanger_isolates/13R.R.ab1")
fwd = primarySeq(fwd)
rev = primarySeq(rev)
rev = reverseComplement(rev)
reads = DNASTringSet(c(as.character(fwd), as.character(rev)))
names(reads) = c('fwd', 'rev')
merged.reads = merge.reads(reads)

## [1] "Aligning reads"
## [1] "Calling consensus sequence"
## [1] "Calculating differences between reads and consensus"

names(merged.reads)

## [1] "consensus"           "alignment"
## [3] "differences"          "distance.matrix"
## [5] "dendrogram"          "indels"
## [7] "stop.codons"         "secondary.peak.columns"

merged.reads

## $consensus
## 1283-letter "DNASTring" instance
## seq: GATGCAATCTACAGATTCTGCATACAGGCTACTA...TCGCCTACATGAAGTNCGGAATCGCTAGTAAT
C
##
## $alignment
## A DNASTringSet instance of length 3
##   width seq                                     names
## [1] 1283 GATGCAATCTACAGATTCTGC...----- fwd
## [2] 1283 -----...GTNCGGAATCGCTAGTAATC rev
## [3] 1283 GATGCAATCTACAGATTCTGC...GTNCGGAATCGCTAGTAATC consensus
##
## $differences
##   name pairwise.diffs.to.consensus unused.chars
## 1 fwd                60                0
## 2 rev                67                0
##
## $distance.matrix
##           fwd      rev
## fwd 0.0000000 0.2491807
## rev 0.2491807 0.0000000
## attr(,"correction")
## [1] "Jukes-Cantor"
##

```

```

## $dendrogram
## 'dendrogram' with 2 branches and 2 members total, at height 0.1245903
##
## $indels
## NULL
##
## $stop.codons
## NULL
##
## $secondary.peak.columns
## NULL
##
## attr(,"class")
## [1] "merged.read"

merged.reads$consensus

## 1283-letter "DNASTring" instance
## seq: GATGCAATCTACAGATTCTGCATACAGGCTACTA...TCGCCTACATGAAGTNCGGAATCGCTAGTAAT
C

BrowseSeqs(merged.reads$alignment)
merged.reads$secondary.peak.columns

## NULL

merged.reads$differences

## name pairwise.diffs.to.consensus unused.chars
## 1 fwd 60 0
## 2 rev 67 0

write.dna(merged.reads, file="Sanger_isolates/13.fasta", format='fasta', nbc0l =
-1, colsep = "", colw = 1000000)

fwd = readsangerseq("Sanger_isolates/30F.F.ab1")
rev = readsangerseq("Sanger_isolates/30R.R.ab1")
fwd = primarySeq(fwd)
rev = primarySeq(rev)
rev = reverseComplement(rev)
reads = DNASTringSet(c(as.character(fwd), as.character(rev)))
names(reads) = c('fwd', 'rev')
merged.reads = merge.reads(reads)

## [1] "Aligning reads"
## [1] "Calling consensus sequence"
## [1] "Calculating differences between reads and consensus"

names(merged.reads)

## [1] "consensus" "alignment"
## [3] "differences" "distance.matrix"

```

```

## [5] "dendrogram"          "indels"
## [7] "stop.codons"           "secondary.peak.columns"

merged.reads

## $consensus
## 1285-letter "DNASTring" instance
## seq: GGCAGCTTGCTGCTTTGCTGACGAGTGGCGGACG...CGACTCCATGAAGTCGGAATCGCTGTAATCGT
N
##
## $alignment
## A DNASTringSet instance of length 3
## width seq names
## [1] 1285 GGCAGCTTGCTGCTTTGCTGA...----- fwd
## [2] 1285 -----...TCGGAATCGCTGTAATCGTN rev
## [3] 1285 GGCAGCTTGCTGCTTTGCTGA...TCGGAATCGCTGTAATCGTN consensus
##
## $differences
## name pairwise.diffs.to.consensus unused.chars
## 1 fwd 40 0
## 2 rev 35 0
##
## $distance.matrix
## fwd rev
## fwd 0.00000000 0.06081996
## rev 0.06081996 0.00000000
## attr(,"correction")
## [1] "Jukes-Cantor"
##
## $dendrogram
## 'dendrogram' with 2 branches and 2 members total, at height 0.03040998
##
## $indels
## NULL
##
## $stop.codons
## NULL
##
## $secondary.peak.columns
## NULL
##
## attr(,"class")
## [1] "merged.read"

merged.reads$consensus

## 1285-letter "DNASTring" instance
## seq: GGCAGCTTGCTGCTTTGCTGACGAGTGGCGGACG...CGACTCCATGAAGTCGGAATCGCTGTAATCGT
N

```

```

BrowseSeqs(merged.reads$alignment)
merged.reads$secondary.peak.columns

## NULL

merged.reads$differences

##   name pairwise.diffs.to.consensus unused.chars
## 1 fwd                               40          0
## 2 rev                               35          0

write.dna(merged.reads, file="Sanger_isolates/30.fasta", format='fasta', nbc1 =
-1, colsep = "", colw = 1000000)

fwd = readsangerseq("Sanger_isolates/95F.F.ab1")
rev = readsangerseq("Sanger_isolates/95R.R.ab1")
fwd = primarySeq(fwd)
rev = primarySeq(rev)
rev = reverseComplement(rev)
reads = DNASTringSet(c(as.character(fwd), as.character(rev)))
names(reads) = c('fwd', 'rev')
merged.reads = merge.reads(reads)

## [1] "Aligning reads"
## [1] "Calling consensus sequence"
## [1] "Calculating differences between reads and consensus"

names(merged.reads)

## [1] "consensus"           "alignment"
## [3] "differences"          "distance.matrix"
## [5] "dendrogram"           "indels"
## [7] "stop.codons"          "secondary.peak.columns"

merged.reads

## $consensus
## 1289-letter "DNASTring" instance
## seq: NNGCTCTTATGAGGTTAGCGGCGGACGGGTGAGT...TACATGAAGCTGGAATCGCTAGTAATCGCGAT
C
##
## $alignment
## A DNASTringSet instance of length 3
##   width seq                                     names
## [1] 1289 NNGCTCTTATGAGGTTAGCGG...----- fwd
## [2] 1289 -----...AATCGCTAGTAATCGCGATC rev
## [3] 1289 NNGCTCTTATGAGGTTAGCGG...AATCGCTAGTAATCGCGATC consensus
##
## $differences
##   name pairwise.diffs.to.consensus unused.chars
## 1 fwd                               66          0
## 2 rev                               62          0

```



```

##
## $distance.matrix
##      fwd      rev
## fwd 0.00000000 0.09751936
## rev 0.09751936 0.00000000
## attr("correction")
## [1] "Jukes-Cantor"
##
## $dendrogram
## 'dendrogram' with 2 branches and 2 members total, at height 0.04875968
##
## $indels
## NULL
##
## $stop.codons
## NULL
##
## $secondary.peak.columns
## NULL
##
## attr("class")
## [1] "merged.read"

merged.reads$consensus

## 1289-letter "DNASTring" instance
## seq: NNGCTCTTATGAGGTTAGCGCGGACGGGTGAGT...TACATGAAGCTGGAATCGCTAGTAATCGCGAT
C

BrowseSeqs(merged.reads$alignment)
merged.reads$secondary.peak.columns

## NULL

merged.reads$differences

## name pairwise.diffs.to.consensus unused.chars
## 1 fwd 66 0
## 2 rev 62 0

write.dna(merged.reads, file="Sanger_isolates/95.fasta", format='fasta', nbc0l =
-1, colsep = "", colw = 1000000)

fwd = readsangerseq("Sanger_isolates/98F.F.ab1")
rev = readsangerseq("Sanger_isolates/98R.R.ab1")
fwd = primarySeq(fwd)
rev = primarySeq(rev)
rev = reverseComplement(rev)
reads = DNASTringSet(c(as.character(fwd), as.character(rev)))
names(reads) = c('fwd', 'rev')
merged.reads = merge.reads(reads)

```

```

## [1] "Aligning reads"
## [1] "Calling consensus sequence"
## [1] "Calculating differences between reads and consensus"

names(merged.reads)

## [1] "consensus"          "alignment"
## [3] "differences"        "distance.matrix"
## [5] "dendrogram"        "indels"
## [7] "stop.codons"       "secondary.peak.columns"

merged.reads

## $consensus
## 737-letter "DNASTring" instance
## seq: NNAGCGAGKRRCSWCSGCKGMKTRRSWSCSKGKAG...CWYSMMKWCSGARTYSCWTGYAMKCKMGRTYA
G
##
## $alignment
## A DNASTringSet instance of length 3
## width seq names
## [1] 737 NNAGCGAGTGGCGACGGCTGA...NCCCATGCACGCTAGGTT-- fwd
## [2] 737 -----GGAACCTCCGCGGC...TTGCTNGTAATCGCGATCAG rev
## [3] 737 NNAGCGAGKRRCSWCSGCKGM...TYSJWTGYAMKCKMGRTYAG consensus
##
## $differences
## name pairwise.diffs.to.consensus unused.chars
## 1 fwd 399 0
## 2 rev 394 0
##
## $distance.matrix
## fwd rev
## fwd 0.000000 1.029787
## rev 1.029787 0.000000
## attr(,"correction")
## [1] "Jukes-Cantor"
##
## $dendrogram
## 'dendrogram' with 2 branches and 2 members total, at height 0.5148934
##
## $indels
## NULL
##
## $stop.codons
## NULL
##
## $secondary.peak.columns
## NULL
##
## attr(,"class")
## [1] "merged.read"

```

```

merged.reads$consensus

## 737-letter "DNASTring" instance
## seq: NNAGCGAGKRRCSWCSGCKGMKTRRSWCSKKGAG...CWYSMMKWCSGARTYSCWTGYAMKCKMGRTYA
G

BrowseSeqs(merged.reads$alignment)
merged.reads$secondary.peak.columns

## NULL

merged.reads$differences

## name pairwise.diffs.to.consensus unused.chars
## 1 fwd 399 0
## 2 rev 394 0

write.dna(merged.reads, file="Sanger_isolates/98.fasta", format='fasta', nbc1 =
-1, colsep = "", colw = 1000000)

fwd = readsangerseq("Sanger_isolates/137F.F.ab1")
rev = readsangerseq("Sanger_isolates/137R.R.ab1")
fwd = primarySeq(fwd)
rev = primarySeq(rev)
rev = reverseComplement(rev)
reads = DNASTringSet(c(as.character(fwd), as.character(rev)))
names(reads) = c('fwd', 'rev')
merged.reads = merge.reads(reads)

## [1] "Aligning reads"
## [1] "Calling consensus sequence"
## [1] "Calculating differences between reads and consensus"

names(merged.reads)

## [1] "consensus" "alignment"
## [3] "differences" "distance.matrix"
## [5] "dendrogram" "indels"
## [7] "stop.codons" "secondary.peak.columns"

merged.reads

## $consensus
## 1276-letter "DNASTring" instance
## seq: GTTGGAGAGTTGCGAACGGGTGAGTAACGCGTAG...ATGAAGTCGGAATCGCTAGTAATCGCGGATCA
G
##
## $alignment
## A DNASTringSet instance of length 3
## width seq names
## [1] 1276 GTTGGAGAGTTGCGAACGGGT...----- fwd
## [2] 1276 -----...CGCTAGTAATCGCGGATCAG rev

```

```

## [3] 1276 GTTGGAGAGTTGCGAACGGGT...CGCTAGTAATCGCGGATCAG consensus
##
## $differences
##   name pairwise.diffs.to.consensus unused.chars
## 1 fwd                                6           0
## 2 rev                                6           0
##
## $distance.matrix
##           fwd           rev
## fwd 0.00000000 0.03922462
## rev 0.03922462 0.00000000
## attr(,"correction")
## [1] "Jukes-Cantor"
##
## $dendrogram
## 'dendrogram' with 2 branches and 2 members total, at height 0.01961231
##
## $indels
## NULL
##
## $stop.codons
## NULL
##
## $secondary.peak.columns
## NULL
##
## attr(,"class")
## [1] "merged.read"

merged.reads$consensus

## 1276-letter "DNASTring" instance
## seq: GTTGGAGAGTTGCGAACGGGTGAGTAACGCGTAG...ATGAAGTCGGAATCGCTAGTAATCGCGGATCA
G

BrowseSeqs(merged.reads$alignment)
merged.reads$secondary.peak.columns

## NULL

merged.reads$differences

##   name pairwise.diffs.to.consensus unused.chars
## 1 fwd                                6           0
## 2 rev                                6           0

write.dna(merged.reads,file="Sanger_isolates/137.fasta",format='fasta',nbcol
= -1, colsep = "", colw = 1000000)

fwd = readsangerseq("Sanger_isolates/138F.F.ab1")
rev = readsangerseq("Sanger_isolates/138R.R.ab1")

```

```

fwd = primarySeq(fwd)
rev = primarySeq(rev)
rev = reverseComplement(rev)
reads = DNASTringSet(c(as.character(fwd), as.character(rev)))
names(reads) = c('fwd', 'rev')
merged.reads = merge.reads(reads)

## [1] "Aligning reads"
## [1] "Calling consensus sequence"
## [1] "Calculating differences between reads and consensus"

names(merged.reads)

## [1] "consensus"           "alignment"
## [3] "differences"         "distance.matrix"
## [5] "dendrogram"         "indels"
## [7] "stop.codons"        "secondary.peak.columns"

merged.reads

## $consensus
## 1297-letter "DNASTring" instance
## seq: TGACTGCTCTGCCTCCTGATTGAATAATCGTTTG...TGAAGCCGGAATCGCTAGTAATCGCGGATCAG
C
##
## $alignment
## A DNASTringSet instance of length 3
## width seq names
## [1] 1297 TGACTGCTCTGCCTCCTGATT...----- fwd
## [2] 1297 -----...GCTAGTAATCGCGGATCAGC rev
## [3] 1297 TGACTGCTCTGCCTCCTGATT...GCTAGTAATCGCGGATCAGC consensus
##
## $differences
## name pairwise.diffs.to.consensus unused.chars
## 1 fwd 59 0
## 2 rev 59 0
##
## $distance.matrix
## fwd rev
## fwd 0.0000000 0.2335444
## rev 0.2335444 0.0000000
## attr(,"correction")
## [1] "Jukes-Cantor"
##
## $dendrogram
## 'dendrogram' with 2 branches and 2 members total, at height 0.1167722
##
## $indels
## NULL
##
## $stop.codons

```

```

## NULL
##
## $secondary.peak.columns
## NULL
##
## attr(,"class")
## [1] "merged.read"

merged.reads$consensus

## 1297-letter "DNAString" instance
## seq: TGACTGCTCTGCCTCCTGATTGAATAATCGTTTG...TGAAGCCGGAATCGCTAGTAATCGCGGATCAG
C

BrowseSeqs(merged.reads$alignment)
merged.reads$secondary.peak.columns

## NULL

merged.reads$differences

## name pairwise.diffs.to.consensus unused.chars
## 1 fwd 59 0
## 2 rev 59 0

write.dna(merged.reads,file="Sanger_isolates/138.fasta",format='fasta',nbc
ol = -1, colsep = "", colw = 1000000)

fwd = readsangerseq("Sanger_isolates/152F.F.ab1")
rev = readsangerseq("Sanger_isolates/152R.R.ab1")
fwd = primarySeq(fwd)
rev = primarySeq(rev)
rev = reverseComplement(rev)
reads = DNAStringSet(c(as.character(fwd), as.character(rev)))
names(reads) = c('fwd', 'rev')
merged.reads = merge.reads(reads)

## [1] "Aligning reads"
## [1] "Calling consensus sequence"
## [1] "Calculating differences between reads and consensus"

names(merged.reads)

## [1] "consensus" "alignment"
## [3] "differences" "distance.matrix"
## [5] "dendrogram" "indels"
## [7] "stop.codons" "secondary.peak.columns"

merged.reads

## $consensus
## 1288-letter "DNAString" instance
## seq: GCTCACCGAAAGAGGAGTGGCGAACGGGTGAGT...TGAAGCCGGAATCGCTAGTAATCGCGGATCAG

```

```

C
##
## $alignment
##   A DNASTringSet instance of length 3
##     width seq                                     names
## [1] 1288 GCTCACCGGAAAGAGGAGTGG...----- fwd
## [2] 1288 -----...GCTAGTAATCGCGGATCAGC rev
## [3] 1288 GCTCACCGGAAAGAGGAGTGG...GCTAGTAATCGCGGATCAGC consensus
##
## $differences
##   name pairwise.diffs.to.consensus unused.chars
## 1 fwd                               8           0
## 2 rev                               8           0
##
## $distance.matrix
##           fwd           rev
## fwd 0.00000000 0.04859999
## rev 0.04859999 0.00000000
## attr(,"correction")
## [1] "Jukes-Cantor"
##
## $dendrogram
## 'dendrogram' with 2 branches and 2 members total, at height 0.0243
##
## $indels
## NULL
##
## $stop.codons
## NULL
##
## $secondary.peak.columns
## NULL
##
## attr(,"class")
## [1] "merged.read"

merged.reads$consensus

## 1288-letter "DNASTring" instance
## seq: GCTCACCGGAAAGAGGAGTGGCGAACGGGTGAGT...TGAAGCCGGAATCGCTAGTAATCGCGGATCAG
C

BrowseSeqs(merged.reads$alignment)
merged.reads$secondary.peak.columns

## NULL

merged.reads$differences

```

```

## name pairwise.diffs.to.consensus unused.chars
## 1 fwd 8 0
## 2 rev 8 0

write.dna(merged.reads, file="Sanger_isolates/152.fasta", format='fasta', nbc
= -1, colsep = "", colw = 1000000)

fwd = readsangerseq("Sanger_isolates/142F.F.ab1")
rev = readsangerseq("Sanger_isolates/142R.R.ab1")
fwd = primarySeq(fwd)
rev = primarySeq(rev)
rev = reverseComplement(rev)
reads = DNASTringSet(c(as.character(fwd), as.character(rev)))
names(reads) = c('fwd', 'rev')
merged.reads = merge.reads(reads)

## [1] "Aligning reads"
## [1] "Calling consensus sequence"
## [1] "Calculating differences between reads and consensus"

names(merged.reads)

## [1] "consensus" "alignment"
## [3] "differences" "distance.matrix"
## [5] "dendrogram" "indels"
## [7] "stop.codons" "secondary.peak.columns"

merged.reads

## $consensus
## 733-letter "DNASTring" instance
## seq: AGTCTTCTAGTAGMRGSKTRSTRSWTCGCYSASG...RAWTCGSYRWAATSGTRWWMAGATTGAACGCT
C
##
## $alignment
## A DNASTringSet instance of length 3
## width seq names
## [1] 733 -----AGCAGCTTGC...GGTATAAAGATTGAACGCTC fwd
## [2] 733 AGTCTTCTAGTAGAGGGGTAG...CGTGATCAG----- rev
## [3] 733 AGTCTTCTAGTAGMRGSKTRS...SGTRWWMAGATTGAACGCTC consensus
##
## $differences
## name pairwise.diffs.to.consensus unused.chars
## 1 fwd 367 0
## 2 rev 367 0
##
## $distance.matrix
## fwd rev
## fwd 0.000000 1.011149
## rev 1.011149 0.000000
## attr(,"correction")

```



```

## [1] "Jukes-Cantor"
##
## $dendrogram
## 'dendrogram' with 2 branches and 2 members total, at height 0.5055747
##
## $indels
## NULL
##
## $stop.codons
## NULL
##
## $secondary.peak.columns
## NULL
##
## attr(,"class")
## [1] "merged.read"

merged.reads$consensus

## 733-letter "DNASTring" instance
## seq: AGTCTTCTAGTAGMRGSKTRSTRSWTCGCYSASG...RAWTCGSYRWAATSGTRWWMAGATTGAACGCT
C

BrowseSeqs(merged.reads$alignment)
merged.reads$secondary.peak.columns

## NULL

merged.reads$differences

## name pairwise.diffs.to.consensus unused.chars
## 1 fwd 367 0
## 2 rev 367 0

write.dna(merged.reads, file="Sanger_isolates/142.fasta", format='fasta', nbc1
= -1, colsep = "", colw = 1000000)

fwd = readsangerseq("Sanger_isolates/184F.F.ab1")
rev = readsangerseq("Sanger_isolates/184R.R.ab1")
fwd = primarySeq(fwd)
rev = primarySeq(rev)
rev = reverseComplement(rev)
reads = DNASTringSet(c(as.character(fwd), as.character(rev)))
names(reads) = c('fwd', 'rev')
merged.reads = merge.reads(reads)

## [1] "Aligning reads"
## [1] "Calling consensus sequence"
## [1] "Calculating differences between reads and consensus"

names(merged.reads)

```

```

## [1] "consensus"           "alignment"
## [3] "differences"            "distance.matrix"
## [5] "dendrogram"           "indels"
## [7] "stop.codons"          "secondary.peak.columns"

merged.reads

## $consensus
## 1281-letter "DNASTring" instance
## seq: GCTTGACTGAGATGGCGACCGGCGCACGGGTGA...GTGAAGCTGGATTCGCTAGTAATCGCGCATCA
G
##
## $alignment
## A DNASTringSet instance of length 3
##      width seq                      names
## [1] 1281 GCTTGACTGAGATGGCGACC...----- fwd
## [2] 1281 -----...CGCTAGTAATCGCGCATCAG rev
## [3] 1281 GCTTGACTGAGATGGCGACC...CGCTAGTAATCGCGCATCAG consensus
##
## $differences
##      name pairwise.diffs.to.consensus unused.chars
## 1 fwd                      114          0
## 2 rev                      114          0
##
## $distance.matrix
##           fwd      rev
## fwd 0.0000000 0.1589253
## rev 0.1589253 0.0000000
## attr(,"correction")
## [1] "Jukes-Cantor"
##
## $dendrogram
## 'dendrogram' with 2 branches and 2 members total, at height 0.07946267
##
## $indels
## NULL
##
## $stop.codons
## NULL
##
## $secondary.peak.columns
## NULL
##
## attr(,"class")
## [1] "merged.read"

merged.reads$consensus

## 1281-letter "DNASTring" instance
## seq: GCTTGACTGAGATGGCGACCGGCGCACGGGTGA...GTGAAGCTGGATTCGCTAGTAATCGCGCATCA
G

```

```

BrowseSeqs(merged.reads$alignment)
merged.reads$secondary.peak.columns

## NULL

merged.reads$differences

##   name pairwise.diffs.to.consensus unused.chars
## 1  fwd                               114         0
## 2  rev                               114         0

write.dna(merged.reads, file="Sanger_isolates/184.fasta", format='fasta', nbc
= -1, colsep = "", colw = 1000000)

fwd = readsangerseq("Sanger_isolates/185F.F.ab1")
rev = readsangerseq("Sanger_isolates/185R.R.ab1")
fwd = primarySeq(fwd)
rev = primarySeq(rev)
rev = reverseComplement(rev)
reads = DNASTringSet(c(as.character(fwd), as.character(rev)))
names(reads) = c('fwd', 'rev')
merged.reads = merge.reads(reads)

## [1] "Aligning reads"
## [1] "Calling consensus sequence"
## [1] "Calculating differences between reads and consensus"

names(merged.reads)

## [1] "consensus"           "alignment"
## [3] "differences"          "distance.matrix"
## [5] "dendrogram"           "indels"
## [7] "stop.codons"          "secondary.peak.columns"

merged.reads

## $consensus
## 1286-letter "DNASTring" instance
## seq: GCACTGAGATGGCGACCGGCGCACGGGTGAGTAA...CGTGAAGCTGGATTCGCTAGTAATCGCGCATC
A
##
## $alignment
## A DNASTringSet instance of length 3
##   width seq                                     names
## [1] 1286 GCACTGAGATGGCGACCGGCG...----- fwd
## [2] 1286 -----...TCGCTAGTAATCGCGCATCA rev
## [3] 1286 GCACTGAGATGGCGACCGGCG...TCGCTAGTAATCGCGCATCA consensus
##
## $differences
##   name pairwise.diffs.to.consensus unused.chars
## 1  fwd                               45         0
## 2  rev                               45         0

```

```

##
## $distance.matrix
##      fwd      rev
## fwd 0.00000000 0.07544043
## rev 0.07544043 0.00000000
## attr("correction")
## [1] "Jukes-Cantor"
##
## $dendrogram
## 'dendrogram' with 2 branches and 2 members total, at height 0.03772021
##
## $indels
## NULL
##
## $stop.codons
## NULL
##
## $secondary.peak.columns
## NULL
##
## attr("class")
## [1] "merged.read"

merged.reads$consensus

## 1286-letter "DNASTring" instance
## seq: GCACTGAGATGGCGACCGGCGCACGGGTGAGTAA...CGTGAAGCTGGATTTCGCTAGTAATCGCGCATC
A

BrowseSeqs(merged.reads$alignment)
merged.reads$secondary.peak.columns

## NULL

merged.reads$differences

## name pairwise.diffs.to.consensus unused.chars
## 1 fwd 45 0
## 2 rev 45 0

write.dna(merged.reads, file="Sanger_isolates/185.fasta", format='fasta', nbc
= -1, colsep = "", colw = 1000000)

fwd = readsangerseq("Sanger_isolates/186F.F.ab1")
rev = readsangerseq("Sanger_isolates/186R.R.ab1")
fwd = primarySeq(fwd)
rev = primarySeq(rev)
rev = reverseComplement(rev)
reads = DNASTringSet(c(as.character(fwd), as.character(rev)))
names(reads) = c('fwd', 'rev')
merged.reads = merge.reads(reads)

```

```

## [1] "Aligning reads"
## [1] "Calling consensus sequence"
## [1] "Calculating differences between reads and consensus"

names(merged.reads)

## [1] "consensus"          "alignment"
## [3] "differences"         "distance.matrix"
## [5] "dendrogram"         "indels"
## [7] "stop.codons"        "secondary.peak.columns"

merged.reads

## $consensus
## 1270-letter "DNASTring" instance
## seq: GCAACTGAGATGGCGACCGGCGCACGGGTGAGTA...CGTGAAGCTGGATTTCGCTAGTAATCGCGCATC
A
##
## $alignment
## A DNASTringSet instance of length 3
## width seq names
## [1] 1270 GCAACTGAGATGGCGACCGGC...----- fwd
## [2] 1270 -----...TCGCTAGTAATCGCGCATCA rev
## [3] 1270 GCAACTGAGATGGCGACCGGC...TCGCTAGTAATCGCGCATCA consensus
##
## $differences
## name pairwise.diffs.to.consensus unused.chars
## 1 fwd 26 0
## 2 rev 26 0
##
## $distance.matrix
## fwd rev
## fwd 0.00000000 0.07729607
## rev 0.07729607 0.00000000
## attr("correction")
## [1] "Jukes-Cantor"
##
## $dendrogram
## 'dendrogram' with 2 branches and 2 members total, at height 0.03864803
##
## $indels
## NULL
##
## $stop.codons
## NULL
##
## $secondary.peak.columns
## NULL
##
## attr("class")
## [1] "merged.read"

```

```
merged.reads$consensus

## 1270-letter "DNASTring" instance
## seq: GCAACTGAGATGGCGACCGGCGCACGGGTGAGTA...CGTGAAGCTGGATTTCGCTAGTAATCGCGCATC
A

BrowseSeqs(merged.reads$alignment)
merged.reads$secondary.peak.columns

## NULL

merged.reads$differences

## name pairwise.diffs.to.consensus unused.chars
## 1 fwd 26 0
## 2 rev 26 0

write.dna(merged.reads, file="Sanger_isolates/186.fasta", format='fasta', nbcol
= -1, colsep = "", colw = 1000000)
```

Most similar OTUs for each Sanger read

A reference alignment was built from the Sanger reads (for each donor separately) applying the `sina` aligner (Pruesse et al. 2012). OTUs were aligned to this reference Sanger alignment in `mothur` (`align.seqs`), yielding a report with the closest Sanger reference for each OTU based on `kmer` searching (Schloss et al. 2009). The report for each donor was loaded into R, version 3.4.2 (2017-09-28) (R Core Team 2016). For each isolate in the report, the top two OTUs with the highest `SearchScores` were selected. A `fasta` file was constructed containing these OTUs.

```
# Donor1
donor1topotusanger <- read.table('DataS13.report', header=TRUE)
donor1topotusanger <- donor1topotusanger[order(donor1topotusanger$TemplateName,
-donor1topotusanger$SearchScore),]
donor1topotusanger <- donor1topotusanger %>% group_by(TemplateName) %>% dplyr::slice(1:2)
# find these otu's in the fasta
donor1fastaotusforfylogeny <- read.table(DataS17.fasta')
donor1fastaotusforfylogeny <- donor1fastaotusforfylogeny[donor1fastaotusforfylogeny$V1%in%donor1topotusanger$QueryName,]
wantedforfylogeny1 <- paste(RDPtax_noprob[donor1fastaotusforfylogeny$V1,]$Genus, donor1fastaotusforfylogeny$V1, sep=" ")
donor1fastaotusforfylogeny$V1 <- paste(RDPtax_noprob[donor1fastaotusforfylogeny$V1,]$Genus, donor1fastaotusforfylogeny$V1, sep="_")
write.csv2(donor1fastaotusforfylogeny, "donor1fastaotusforfylogeny.csv", row.names=FALSE, col.names=FALSE)

# Donor2
donor2topotusanger <- read.table('DataS14.report', header=TRUE)
donor2topotusanger <- donor2topotusanger[order(donor2topotusanger$TemplateName,
-donor2topotusanger$SearchScore),]
```

```

donor2topotusanger <- donor2topotusanger%>%group_by(TemplateName) %>%dplyr::sli
lice(1:2)
donor2fastaotusforfylogeny <- read.table('DataS17.fasta')
donor2fastaotusforfylogeny <- donor2fastaotusforfylogeny[donor2fastaotusforfy
logeny$V1%in%donor2topotusanger$QueryName,]
wantedforfylogeny2 <-paste(RDPtax_noprob[donor2fastaotusforfylogeny$V1,]$Genu
s,donor2fastaotusforfylogeny$V1,sep=" ")
donor2fastaotusforfylogeny$V1 <- paste(RDPtax_noprob[donor2fastaotusforfyloge
ny$V1,]$Genus,donor2fastaotusforfylogeny$V1,sep="_")
write.csv2(donor2fastaotusforfylogeny,"donor2fastaotusforfylogeny.csv",row.na
mes=FALSE,col.names=FALSE)

```

Donor3

```

donor3topotusanger <- read.table('DataS15.report',header=TRUE)
donor3topotusanger <- donor3topotusanger[order(donor3topotusanger$TemplateNam
e,-donor3topotusanger$SearchScore),]
donor3topotusanger <-donor3topotusanger%>%group_by(TemplateName) %>%dplyr::sli
lice(1:2)
donor3fastaotusforfylogeny <- read.table('DataS17.fasta')
donor3fastaotusforfylogeny <- donor3fastaotusforfylogeny[donor3fastaotusforfy
logeny$V1%in%donor3topotusanger$QueryName,]
wantedforfylogeny3 <-paste(RDPtax_noprob[donor3fastaotusforfylogeny$V1,]$Genu
s,donor3fastaotusforfylogeny$V1,sep=" ")
donor3fastaotusforfylogeny$V1 <- paste(RDPtax_noprob[donor3fastaotusforfyloge
ny$V1,]$Genus,donor3fastaotusforfylogeny$V1,sep="_")
write.csv2(donor3fastaotusforfylogeny,"donor3fastaotusforfylogeny.csv",row.na
mes=FALSE,col.names=FALSE)

```

Donor4

```

donor4topotusanger <- read.table('DataS16.report',header=TRUE)
donor4topotusanger <- donor4topotusanger[order(donor4topotusanger$TemplateNam
e,-donor4topotusanger$SearchScore),]
donor4topotusanger <-donor4topotusanger%>%group_by(TemplateName) %>%dplyr::sli
lice(1:2)
donor4fastaotusforfylogeny <- read.table('DataS18.fasta')
donor4fastaotusforfylogeny <- donor4fastaotusforfylogeny[donor4fastaotusforfy
logeny$V1%in%donor4topotusanger$QueryName,]
wantedforfylogeny4 <-paste(RDPtax4_noprob[donor4fastaotusforfylogeny$V1,]$Gen
us,donor4fastaotusforfylogeny$V1,sep=" ")
donor4fastaotusforfylogeny$V1 <- paste(RDPtax4_noprob[donor4fastaotusforfylog
eny$V1,]$Genus,donor4fastaotusforfylogeny$V1,sep="_")
write.csv2(donor4fastaotusforfylogeny,"donor4fastaotusforfylogeny.csv",row.na
mes=FALSE,col.names=FALSE)

```

Create metadata for the annotation of iTOL phylogenetic trees

In order to compare the OTUs spanning the V4 region of the 16S rRNA gene with the near full-length Sanger reads, the RAxML implementation of the evolutionary placement algorithm of short reads, as introduced by Berger et al. (2011), was used (Stamatakis 2014). The bootstrap supported maximum likelihood (ML) phylogenetic reference tree was also constructed using RAxML, selecting the General Time Reversible model of nucleotide substitution under the Gamma model of rate heterogeneity (GTRGAMMA) with the parsimony random seed set to 12345. The rapid bootstrap analysis was conducted starting from N=1000 distinct randomized maximum parsimony trees and was followed by a search for the best-scoring ML tree with rapid bootstrap random number seed 123 (Stamatakis 2014). The best scoring ML tree with the OTU short read insertions was visualized in iTOL (Letunic & Bork 2016). Metadata for the annotation of the iTOL trees is created in the chunks below.

```
currentotuname1 <- donor1fastaotusforfylogeny$V1
RAXMLotuname1 <- paste("QUERY___",currentotuname1,sep="")
otuname1df <- cbind(currentname1=RAXMLotuname1,wantedname1=wantedforfylogeny1
)

wantedsangername1 <- splittax(read.table('DataS19.taxonomy'),'seqs')
wantedsangername1$Genus <- mapvalues(wantedsangername1$Genus,from=c("Enterobacteriaceae_unclassified","Erysipelotrichaceae_unclassified","Lachnospiraceae_unclassified","Lactobacillales_unclassified","Actinobacteria_unclassified","Burkholderiales_unclassified","Clostridiales_unclassified"),to=c("Enterobacteriaceae","Erysipelotrichaceae","Lachnospiraceae","Lactobacillales","Actinobacteria","Burkholderiales","Clostridiales"))
currentsangername1 <- as.character(wantedsangername1$id)
wantedsangername1 <- as.character(paste(wantedsangername1$id,wantedsangername1$Genus))
sangername1df <- cbind(currentname1=currentsangername1,wantedname1=wantedsangername1)

itolname1 <- data.frame(rbind(otuname1df,sangername1df))
itolname1$itolcode <- paste(itolname1$currentname1,",",itolname1$wantedname1,sep="")
itollabel1<- c("LABELS","SEPARATOR COMMA","DATA",itolname1$itolcode)
write.table(itollabel1,"itollabel1.txt",row.names=FALSE,col.names=FALSE,quote=FALSE)

#style labels
itollabelstyle1 <- paste(currentsangername1,",label",",#000000",",bold",",1",sep="")
itollabelstyle1<- c("TREE_COLORS","SEPARATOR COMMA","DATA",itollabelstyle1)
write.table(itollabelstyle1,"itollabelstyle1.txt",row.names=FALSE,col.names=FALSE,quote=FALSE)

# Stacked bar charts of the relative abundance
propbar1 <- subset(shared3mingsingletonwnwnprop*100,select=c(Donor1_high4_L.V4
```



```

,Donor1_low4_L.V4,FS_1_isolationhigh.V4))
propbar1 <- propbar1[rownames(propbar1)%in%donor1topotusanger$QueryName,]
propbar1 <- round(propbar1,2)
propbar1 <- rownames_to_column(propbar1)
propbar1$rowname <- paste(RDPtax_noprob[propbar1$rowname,]$Genus,propbar1$rowname,sep="_")
propbar1$rowname<- paste("QUERY___",propbar1$rowname,sep="")
propbar1$high <- paste(",",propbar1$Donor1_high4_L.V4,sep="")
propbar1$low<- paste(",",propbar1$Donor1_low4_L.V4,sep="")
propbar1$FS <- paste(",",propbar1$FS_1_isolationhigh.V4,sep="")
propbar1 <- propbar1[,-c(2:4)]
propbar1 <- rbind(c("DATA","", "", ""),propbar1)
propbar1 <- rbind(c("DATASET_SCALE","", "10","", "25","", "50"),propbar1)
propbar1 <- rbind(c("ALIGN_FIELDS","", "0","", ""),propbar1)
propbar1 <- rbind(c("LEGEND_TITLE","", "Relative abundance (%)","", ""),propbar1)
propbar1 <- rbind(c("LEGEND_SHAPES","", "1","", "1","", "1"),propbar1)
propbar1 <- rbind(c("LEGEND_COLORS","", "#323C4D","", "#A03D44","", "#98B736"),propbar1)
propbar1 <- rbind(c("LEGEND_LABELS","", "pH6.8","", "pH5.8","", "Fecal slurry"),propbar1)
propbar1 <- rbind(c("WIDTH","", "350","", ""),propbar1)
propbar1 <- rbind(c("FIELD_COLORS","", "#323C4D","", "#A03D44","", "#98B736"),propbar1)
propbar1 <- rbind(c("FIELD_LABELS","", "f1","", "f2","", "f3"),propbar1)
propbar1 <- rbind(c("COLOR","", "#ff0000","", ""),propbar1)
propbar1 <- rbind(c("DATASET_LABEL","", "barchart","", ""),propbar1)
propbar1 <- rbind(c("SEPARATOR","", "COMMA","", ""),propbar1)
propbar1 <- rbind(c("DATASET_MULTIBAR","", "", "", ""),propbar1)
write.table(propbar1, "itolpropbar1.txt", col.names=FALSE, row.names=FALSE, sep=" ", quote=FALSE)

currentotuname2 <- donor2fastaotusforfylogeny$V1
RAXMLotuname2 <- paste("QUERY___",currentotuname2,sep="")
otuname2df <- cbind(currentname2=RAXMLotuname2,wantedname2=wantedforfylogeny2)

wantsangname2 <- splittax(read.table('DataS20.taxonomy'),"seqs")
wantsangname2$Genus <- mapvalues(wantsangname2$Genus,from=c("Enterobacteriaceae_unclassified","Erysipelotrichaceae_unclassified","Lachnospiraceae_unclassified","Lactobacillales_unclassified","Actinobacteria_unclassified","Burkholderiales_unclassified","Clostridiales_unclassified"),to=c("Enterobacteriaceae","Erysipelotrichaceae","Lachnospiraceae","Lactobacillales","Actinobacteria","Burkholderiales","Clostridiales"))
currentsangname2 <- as.character(wantsangname2$id)
wantsangname2 <- as.character(paste(wantsangname2$id,wantsangname2$Genus))
sangname2df <- cbind(currentname2=currentsangname2,wantedname2=wantsangname2)

itolname2 <- data.frame(rbind(otuname2df, sangname2df))

```

```

itolname2$itolcode <- paste(itolname2$currentname2,"",itolname2$wantedname2,
sep="")
itollabel2<- c("LABELS","SEPARATOR COMMA","DATA",itolname2$itolcode)
write.table(itollabel2,"itollabel2.txt",row.names=FALSE,col.names=FALSE,quote
=FALSE)

#style Labels
itollabelstyle2 <- paste(currentsangername2,"label",",#000000",",bold",",1",
sep="")
itollabelstyle2<- c("TREE_COLORS","SEPARATOR COMMA","DATA",itollabelstyle2)
write.table(itollabelstyle2,"itollabelstyle2.txt",row.names=FALSE,col.names=F
ALSE,quote=FALSE)

# Stacked bar charts of the relative abundance
propbar2 <- subset(shared3mingsingletonwnwnprop*100,select=c(Donor2_high4_L.V4
,Donor2_low4_L.V4,FS_2_isolationhigh.V4))
propbar2 <- propbar2[rownames(propbar2)%in%donor2topotusanger$QueryName,]
propbar2 <- round(propbar2,2)
propbar2 <- rownames_to_column(propbar2)
propbar2$rowname <-paste(RDPtax_noprof[propbar2$rowname,]$Genus,propbar2$rowname,sep="_")
propbar2$rowname<- paste("QUERY___",propbar2$rowname,sep="")
propbar2$high <- paste(",",propbar2$Donor2_high4_L.V4,sep="")
propbar2$low<- paste(",",propbar2$Donor2_low4_L.V4,sep="")
propbar2$FS <- paste(",",propbar2$FS_2_isolationhigh.V4,sep="")
propbar2 <- propbar2[,-c(2:4)]
propbar2 <- rbind(c("DATA", "", "", ""),propbar2)
propbar2 <- rbind(c("DATASET_SCALE", "", "5", "", "10", "", "25"),propbar2)
propbar2 <- rbind(c("ALIGN_FIELDS", "", "0", "", ""),propbar2)
propbar2 <- rbind(c("LEGEND_TITLE", "", "Relative abundance (%)", "", ""),propbar2)
propbar2 <- rbind(c("LEGEND_SHAPES", "", "1", "", "1", "", "1"),propbar2)
propbar2 <- rbind(c("LEGEND_COLORS", "", "#323C4D", "", "#A03D44", "", "#98B736"),propbar2)
propbar2 <- rbind(c("LEGEND_LABELS", "", "pH6.8", "", "pH5.8", "", "Fecal slurry"),propbar2)
propbar2 <- rbind(c("WIDTH", "", "350", "", ""),propbar2)
propbar2 <- rbind(c("FIELD_COLORS", "", "#323C4D", "", "#A03D44", "", "#98B736"),propbar2)
propbar2 <- rbind(c("FIELD_LABELS", "", "f1", "", "f2", "", "f3"),propbar2)
propbar2 <- rbind(c("COLOR", "", "#ff0000", "", ""),propbar2)
propbar2 <- rbind(c("DATASET_LABEL", "", "barchart", "", ""),propbar2)
propbar2 <- rbind(c("SEPARATOR", " COMMA", "", ""),propbar2)
propbar2 <- rbind(c("DATASET_MULTIBAR", "", "", ""),propbar2)
write.table(propbar2,"itolpropbar2.txt",col.names=FALSE,row.names=FALSE,sep="
",quote=FALSE)

currenttotuname3 <- donor3fastaotusforfylogeny$V1
RAXMLotuname3 <- paste("QUERY___",currenttotuname3,sep="")
otuname3df <- cbind(currentname3=RAXMLotuname3,wantedname3=wantedforfylogeny3
)

```

```

wantedsangername3 <- splittax(read.table('DataS21.taxonomy'), "seqs")
wantedsangername3$Genus <- mapvalues(wantedsangername3$Genus, from=c("Enterobacteriaceae_unclassified", "Erysipelotrichaceae_unclassified", "Lachnospiraceae_unclassified", "Lactobacillales_unclassified", "Actinobacteria_unclassified", "Burkholderiales_unclassified", "Clostridiales_unclassified"), to=c("Enterobacteriaceae", "Erysipelotrichaceae", "Lachnospiraceae", "Lactobacillales", "Actinobacteria", "Burkholderiales", "Clostridiales"))
currentsangername3 <- as.character(wantedsangername3$id)
wantedsangername3 <- as.character(paste(wantedsangername3$id, wantedsangername3$Genus))
sangername3df <- cbind(currentname3=currentsangername3, wantedname3=wantedsangername3)

itolname3 <- data.frame(rbind(otuname3df, sangername3df))
itolname3$itolcode <- paste(itolname3$currentname3, ",", itolname3$wantedname3, sep="")
itollabel3 <- c("LABELS", "SEPARATOR COMMA", "DATA", itolname3$itolcode)
write.table(itollabel3, "itollabel3.txt", row.names=FALSE, col.names=FALSE, quote=FALSE)

#style labels
itollabelstyle3 <- paste(currentsangername3, "label", "#000000", "bold", "1", sep="")
itollabelstyle3 <- c("TREE_COLORS", "SEPARATOR COMMA", "DATA", itollabelstyle3)
write.table(itollabelstyle3, "itollabelstyle3.txt", row.names=FALSE, col.names=FALSE, quote=FALSE)

# Stacked bar charts of the relative abundance
propbar3 <- subset(shared3misingletonwnwnprop*100, select=c(Donor3_high4_L.V4, Donor3_low4_L.V4, FS_3_isolationhigh.V4))
propbar3 <- propbar3[rownames(propbar3)%in%donor3topotusanger$QueryName, ]
propbar3 <- round(propbar3, 2)
propbar3 <- rownames_to_column(propbar3)
propbar3$rowname <- paste(RDPtax_noprob[propbar3$rowname,]$Genus, propbar3$rowname, sep="_")
propbar3$rowname <- paste("QUERY__", propbar3$rowname, sep="")
propbar3$high <- paste("", propbar3$Donor3_high4_L.V4, sep="")
propbar3$low <- paste("", propbar3$Donor3_low4_L.V4, sep="")
propbar3$FS <- paste("", propbar3$FS_3_isolationhigh.V4, sep="")
propbar3 <- propbar3[, -c(2:4)]
propbar3 <- rbind(c("DATA", "", "", ""), propbar3)
propbar3 <- rbind(c("DATASET_SCALE", "25", "50", "100"), propbar3)
propbar3 <- rbind(c("ALIGN_FIELDS", "0", "", ""), propbar3)
propbar3 <- rbind(c("LEGEND_TITLE", "Relative abundance (%)", "", ""), propbar3)
propbar3 <- rbind(c("LEGEND_SHAPES", "1", "1", "1"), propbar3)
propbar3 <- rbind(c("LEGEND_COLORS", "#323C4D", "#A03D44", "#98B736"), propbar3)
propbar3 <- rbind(c("LEGEND_LABELS", "pH6.8", "pH5.8", "Fecal slurry"), propbar3)

```

```

propbar3 <- rbind(c("WIDTH",",350",",",""),propbar3)
propbar3 <- rbind(c("FIELD_COLORS",",#323C4D",",#A03D44",",#98B736"),propbar3
)
propbar3 <- rbind(c("FIELD_LABELS",",f1",",f2",",f3"),propbar3)
propbar3 <- rbind(c("COLOR",",#ff0000",",",""),propbar3)
propbar3 <- rbind(c("DATASET_LABEL",",barchart",",",""),propbar3)
propbar3 <- rbind(c("SEPARATOR",", COMMA",",",""),propbar3)
propbar3 <- rbind(c("DATASET_MULTIBAR",",","",""),propbar3)
write.table(propbar3,"itolpropbar3.txt",col.names=FALSE,row.names=FALSE,sep="
",quote=FALSE)

currentotuname4 <- donor4fastaotusforfylogeny$V1
RAXMLotuname4 <- paste("QUERY___",currentotuname4,sep="")
otuname4df <- cbind(currentname4=RAXMLotuname4,wantedname4=wantedforfylogeny4
)

wantsangername4 <- splittax(read.table('DataS22.taxonomy'),"seqs")
wantsangername4$Genus <- mapvalues(wantsangername4$Genus,from=c("Enteroba
cteriaceae_unclassified","Erysipelotrichaceae_unclassified","Lachnospiraceae_
unclassified","Lactobacillales_unclassified","Actinobacteria_unclassified","B
urkholderiales_unclassified","Clostridiales_unclassified","Ruminococcaceae_un
classified","Coriobacteriaceae_unclassified","Lactobacillaceae_unclassified",
"Microbacteriaceae_unclassified"),to=c("Enterobacteriaceae","Erysipelotrichac
eae","Lachnospiraceae","Lactobacillales","Actinobacteria","Burkholderiales",
"Clostridiales","Ruminococcaceae","Coriobacteriaceae","Lactobacillaceae","Micr
obacteriaceae"))
currentsangername4 <- as.character(wantsangername4$id)
wantsangername4 <- as.character(paste(wantsangername4$id,wantsangername
4$Genus))
sangername4df <- cbind(currentname4=currentsangername4,wantedname4=wantsang
ername4)

itolname4 <- data.frame(rbind(otuname4df,sangername4df))
itolname4$itolcode <- paste(itolname4$currentname4,",",itolname4$wantedname4,
sep="")
itollabel4<- c("LABELS","SEPARATOR COMMA","DATA",itolname4$itolcode)
write.table(itollabel4,"itollabel4.txt",row.names=FALSE,col.names=FALSE,quote
=FALSE)

#style labels
itollabelstyle4 <- paste(currentsangername4,",label",",#000000",",bold",",1",
sep="")
itollabelstyle4<- c("TREE_COLORS","SEPARATOR COMMA","DATA",itollabelstyle4)
write.table(itollabelstyle4,"itollabelstyle4.txt",row.names=FALSE,col.names=F
ALSE,quote=FALSE)

# Stacked bar charts of the relative abundance
propbar4 <- subset(shared4mingsingletonwnwnprop*100,select=c(Kim8_Donor4_high4
_L,Kim4_Donor4_low4_L,Kim1_Donor4_FS_high))
propbar4 <- propbar4[rownames(propbar4)%in%donor4topotusanger$QueryName,]

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```

propbar4 <- round(propbar4,2)
propbar4 <- rownames_to_column(propbar4)
propbar4$rowname <-paste(RDPtax4_noprob[propbar4$rowname,]$Genus,propbar4$rowname,sep="_")
propbar4$rowname<- paste("QUERY___",propbar4$rowname,sep="")
propbar4$high <- paste(" ",propbar4$Kim8_Donor4_high4_L,sep="")
propbar4$low<- paste(" ",propbar4$Kim4_Donor4_low4_L,sep="")
propbar4$FS <- paste(" ",propbar4$Kim1_Donor4_FS_high,sep="")
propbar4 <- propbar4[, -c(2:4)]
propbar4 <- rbind(c("DATA", "", "", "" ),propbar4)
propbar4 <- rbind(c("DATASET_SCALE", "25", "50", "100"),propbar4)
propbar4 <- rbind(c("ALIGN_FIELDS", "0", "", "" ),propbar4)
propbar4 <- rbind(c("LEGEND_TITLE", "Relative abundance (%)", "", "" ),propbar4)
propbar4 <- rbind(c("LEGEND_SHAPES", "1", "1", "1"),propbar4)
propbar4 <- rbind(c("LEGEND_COLORS", "#323C4D", "#A03D44", "#98B736"),propbar4)
propbar4 <- rbind(c("LEGEND_LABELS", "pH6.8", "pH5.8", "Fecal slurry"),propbar4)
propbar4 <- rbind(c("WIDTH", "350", "", "" ),propbar4)
propbar4 <- rbind(c("FIELD_COLORS", "#323C4D", "#A03D44", "#98B736"),propbar4)
propbar4 <- rbind(c("FIELD_LABELS", "f1", "f2", "f3"),propbar4)
propbar4 <- rbind(c("COLOR", "#ff0000", "", "" ),propbar4)
propbar4 <- rbind(c("DATASET_LABEL", "barchart", "", "" ),propbar4)
propbar4 <- rbind(c("SEPARATOR", " COMMA", "", "" ),propbar4)
propbar4 <- rbind(c("DATASET_MULTIBAR", "", "", "" ),propbar4)
write.table(propbar4, "itolpropbar4.txt", col.names=FALSE, row.names=FALSE, sep=" ", quote=FALSE)

```